OPEN O ACCESS Freely available online

Low Dose Prenatal Alcohol Exposure Does Not Impair Spatial Learning and Memory in Two Tests in Adult and Aged Rats

Carlie L. Cullen¹, Thomas H. J. Burne², Nickolas A. Lavidis¹, Karen M. Moritz¹*

1 School of Biomedical Sciences, The University of Queensland, St. Lucia, Australia, 2 Queensland Brain Institute, The University of Queensland, St. Lucia, Australia

Abstract

Consumption of alcohol during pregnancy can have detrimental impacts on the developing hippocampus, which can lead to deficits in learning and memory function. Although high levels of alcohol exposure can lead to severe deficits, there is a lack of research examining the effects of low levels of exposure. This study used a rat model to determine if prenatal exposure to chronic low dose ethanol would result in deficits in learning and memory performance and if this was associated with morphological changes within the hippocampus. Sprague Dawley rats were fed a liquid diet containing 6% (vol/vol) ethanol (EtOH) or an isocaloric control diet throughout gestation. Male and Female offspring underwent behavioural testing at 8 (Adult) or 15 months (Aged) of age. Brains from these animals were collected for stereological analysis of pyramidal neuron number and dendritic morphology within the CA1 and CA3 regions of the dorsal hippocampus. Prenatal ethanol exposure to Control offspring. There was no effect of prenatal ethanol exposure on pyramidal cells within the dorsal hippocampus. Overall, this study indicates that chronic low dose prenatal ethanol exposure on pyramidal cells within the dorsal hippocampus or impair spatial learning and memory performance.

Citation: Cullen CL, Burne THJ, Lavidis NA, Moritz KM (2014) Low Dose Prenatal Alcohol Exposure Does Not Impair Spatial Learning and Memory in Two Tests in Adult and Aged Rats. PLoS ONE 9(6): e101482. doi:10.1371/journal.pone.0101482

Editor: Jill C. Bettinger, Virginia Commonwealth University, United States of America

Received November 5, 2013; Accepted June 6, 2014; Published June 30, 2014

Copyright: © 2014 Cullen et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by the National Health and Medical Research Council of Australia. CC was the recipient of an Australian Post-graduate award. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: One of the authors of this manuscript, Tom Burne, currently serves as an editor for PLOS ONE. This does not alter the authors' adherence to PLOS ONE Editorial policies and criteria.

* Email: k.moritz1@uq.edu.au

Introduction

The consumption of alcohol during pregnancy can have detrimental neurodevelopmental outcomes for the unborn child [1,2]. The most recognisable and severe effects of prenatal alcohol exposure include craniofacial malformations, growth deficiency, microencephaly and mental retardation characteristic of Foetal Alcohol Syndrome (FAS) [3,4]. However, FAS is the least common effect of prenatal alcohol exposure occurring in only 5% of births to mothers who consumed alcohol during pregnancy and is typically associated with alcoholic mothers prone to heavy drinking [5,6]. There is a large variety of poor neurodevelopmental and cognitive outcomes relating to fetal alcohol exposure that can be grouped under the umbrella term Fetal Alcohol Spectrum Disorders (FASD) [7-9]. While anxiety related disorders, depressive disorders, difficulties in social interaction and engagement and problems with hyperactivity and attention are among some of these poor outcomes [10-15], perhaps the most commonly reported problems relating to fetal alcohol exposure are those associated with learning and memory [14,16-22].

Clinical studies have repeatedly demonstrated a strong association between the consumption of alcohol during gestation and deficits in learning and memory in offspring. Although some research has shown that these deficits can persist into adulthood [17,23], investigations of learning and memory difficulties associated with prenatal alcohol exposure are most commonly conducted during schooling, where these children exhibit poor academic performance often associated with a decreased propensity for learning as well as problems with memory retention [14,20,21,24].

Interestingly, some studies in animals have reported no-effects or even subtle improvements in learning and memory performance following varying doses and periods of prenatal alcohol exposure [25–27]. However, the majority of animal models of FASD report deficits in learning and memory performance [2,28– 32]. While some studies show that these impairments persist through life [33,34] others demonstrate recovery of learning and memory capabilities in adult animals [35].

Although there is some evidence for a dose dependant relationship between the severity of these deficits and the amount of alcohol consumed [36,37], the majority of animal research has been conducted using offspring exposed to levels of alcohol within the moderate to high range. Very few studies have investigated the impact of relatively low levels of alcohol exposure in utero on learning and memory performance in the offspring. One exception to this is a study conducted by Savage and colleagues [38], who demonstrated spatial learning deficits in 7 month old male rat offspring prenatally exposed to peak maternal blood ethanol concentrations of 30–80 mg/dl. Moreover, Savage and colleagues [38] also demonstrated decreases in evoked release of excitatory

neurotransmitters within the dorsal hippocampus of female offspring.

Learning and memory function is primarily governed by the hippocampus, a region shown to be particularly vulnerable to the teratogenic effects of alcohol [39]. Exposure to moderate to high levels of alcohol during development has been shown to reduce pyramidal cell number in the CA1 and CA3 regions of the hippocampus [40]. Alcohol exposure at the moderate to high levels during gestation has also been shown to induce marked alterations in the cytoarchitecture of pyramidal cell dendrites within the hippocampus as well as impaired long term potentiation (LTP) induction in CA1 pyramidal cells [29,41–43]. Although there is some evidence of reduced functionality of pyramidal cell synapses within the hippocampus following exposure to relatively small amounts of alcohol [44], to our knowledge no study has looked at the dendritic morphology of pyramidal cells in adult and aged animals following low dose prenatal alcohol exposure.

We have recently developed a model of low dose ethanol exposure [45] and shown that exposure to this low dose is sufficient to induce an anxiety-like phenotype in adult and aged rats [46]. In the current study, we utilised the same model to determine the long term impacts of chronic prenatal exposure to low dose ethanol on hippocampal-dependent learning and memory tasks, and on the morphology of pyramidal cells within the dorsal hippocampus.

Materials and Methods

Ethics Statement

All animal procedures were approved by the Animal Welfare Unit of The University of Queensland and follow the code of practice for the care and use of animals for scientific purposes.

Animals and Housing

All animals were housed under temperature and humidity controlled conditions under a 12 hr light cycle (lights on at 12:00). Female nulliparous Sprague Dawley (University of Queensland Biological Resources, St. Lucia, QLD, AUS) rats were trialled on a liquid diet modelled from the commercially available Leiber DeCarli diet [47] then time mated from 8 weeks of age. Successful mating was confirmed by the presence of seminal plugs and this was marked as embryonic day (E) 1. The pregnant females were then housed singly and randomly assigned to receive a liquid diet containing 6% (vol/vol) ethanol, 15% ethanol derived calories (EDC) (n = 15), or an isocaloric control diet (n = 16) ad libitum for 21 hours a day for the duration of pregnancy (E1-22/23). Water was offered for the remaining 3 hours a day (09:00-12:00) and water consumption over this period of time was recorded. Each dam was offered 80 mL of fresh diet each day. Both experimental diets contained 24.4% w/v Sustagen (Hospital Formula; Mead Johnson Nutritionals, Auckland, New Zealand), 1.04% v/v sunflower oil, 25 mg selenium (Selemite B; Blackmores, Warriewood, NSW, Australia), 73% v/v reduced fat milk, 50 mM copper sulfate (Sigma Aldrich, Sydney, NSW, Australia), 199 mM ferric citrate (Sigma Aldrich) and 303 mM manganese sulfate (Sigma Aldrich). The control diet also contained 19.6% w/v corn starch. The ethanol diet (EtOH) contained 6% v/v absolute ethanol as well as a reduced amount of corn starch (11.5% w/v) to account for the additional calories provided by the ethanol. This ensured the diets were approximately isocaloric with a total of 499.2 calories (2096.6 kJ) in the control diet and 509.4 calories (2139.5 kJ) in the EtOH diet. The dams were weighed daily at the time of diet removal but were otherwise left undisturbed. At parturition the experimental diet was removed and *ad libitum* access to standard rat chow and water was then maintained.

Offspring

As previously reported the rat pups were weighed every 3 days from postnatal day (PN) 1 to PN28 when the pups were weaned, separated by sex and housed in littermate groups with *ad libitum* access to standard rat chow and water until behavioural testing [45]. Animals were tested in one of two age groups, 8–10 months (Adult) or 15–18 months (Aged) with no more than two males or two female littermates were used at any age to minimise any effect of litter bias. The number of animals included in the analysis for individual tests are indicated in the figure legends.

Behavioural Testing

All behavioural testing was carried out during the dark phase of the light cycle. Prior to behavioural testing animals were moved to a testing room, in their home cages, one hour after the light cycle change and left to habituate to the room for one hour. Dim lights (~ 20 lux) and a small colour camera were situated above the testing arena. Extra-maze cues (i.e. vertical and horizontal bars) were visible around the room. All trials were video recorded and animal movement tracked using automated software (Ethovision ver. 5, Noldus, Netherlands).

Y maze

Short term spatial memory performance was assessed using a Y maze task [48,49] with a black, PVC plastic maze consisting of three identical arms (50 cm×10 cm×30 cm) randomly designated as start, familiar or novel. The task consisted of two trials, the acquisition phase and the test phase. During the acquisition phase entry into the novel arm was blocked and rats were placed into the start arm and left to explore the start and familiar arms freely for 15 minutes. After the acquisition phase rats were returned to their home cage for 30 minutes. The rats were then placed back in the maze for the test phase, in which the novel arm was unblocked, and were allowed to explore the entire maze for 5 minutes. To prevent odour cues the bottom of the maze was lined with soiled animal bedding which was mixed between each trial. Spatial memory performance was measured as the number of entries into and time spent in the novel arm, calculated as a percentage of total arm entries and total time in the maze respectively.

Water Maze

The water maze task employed was an adaptation of the hidden escape paradigm described by Morris [50,51]. A black platform (10 cm d×30 cm h) was placed in an off centre position inside a circular grey acrylic tank (110 cm d×50 cm h) containing warm water $(24\pm1^{\circ}C)$ filled to a level 1 cm above the platform. To ensure the platform was hidden the water was made opaque using a small amount of black non-toxic water based poster paint. The paradigm consisted of 4 trials per day over 4 consecutive days in which the rats were placed at one of three different start positions for each trial and given 60 seconds to locate the hidden platform. The rat was left on the platform for 10 seconds before being picked up towel dried and returned to its home cage for a 10 minute ITI. If after 60 seconds the platform was not located the rat was guided to the platform and left there for 10 seconds. The latency to locate the hidden platform was recorded for each trial as a measure of spatial learning performance. On the fifth day the rats were given a single 60 second probe trial in which the platform was removed. To assess spatial memory performance the water maze was divided into four quadrants within the tracking software (Ethovision ver. 5, Noldus, Netherlands). The quadrant in which the platform was previously situated was defined as the target quadrant and the remaining quadrants defined as opposite and adjacent. The position where the platform had been located was also outlined and the number of times this region was crossed as well as the percentage time spent in the target quadrant was analysed as a measure of spatial memory performance.

Tissue Collection

After behavioural testing the rats were euthanized with an overdose of sodium pentobarbital (Lethabarb; 0.1 ml/kg body-weight). Animals were then transcardially perfused with heparinised 0.2 M Phosphate Buffered Saline (PBS) and fixed with 4% paraformaldehyde (Sigma Aldrich) in PBS solution. The fixed brains were then collected and stored in 0.4% PFA for histological processing.

Histology

Brain tissue was prepared and analysed as described previously [46]. Power calculations indicated that a minimum of five subjects per group would be required to detect a significant difference in cell number between treatment groups at 80% power. Therefore, a minimum of n = 5 brains were used for all histological analysis. In brief, for stereological investigations 50 µm coronal sections were cut from n = 6 brains per sex, per treatment from both age groups and stained using 0.1% Cresyl Violet. Unbiased stereological analysis was performed using StereoInvestigator (ver. 9, Mbf Biosciences) software. Pyramidal cell number within the dorsal hippocampus (bregma -2.28 to -3.64; [52]) was determined by the optical fractionator method using a grid size of 150×150 µm and 50×50 µm counting frames [53]. Pyramidal neurons within the stratum pyramidale were counted in a total of six slices with a three slice interval between. Regional hippocampal volume was determined using the Cavalieri estimator. Regional numerical cell density was calculated per 1000 μm^3 (total # cells/ volume*1000).

Investigation of dendritic morphology was performed by submerging agarose gel (4%) blocked hemispheres Golgi Cox Solution (5% Potassium dichromate, 5% Potassium chromate, 5% Mercuric chloride) for 33 days, changing the solution weekly. These Golgi impregnated brains were then coronally sectioned at 300 µm, processed as per [54] and mounted on gelatin coated slides. Apical dendrites for six pyramidal cells per animal, per region were traced using NeuroLucida (ver. 9, Mbf Biosciences) and branching morphology determined using branched structure analysis. Neurons within the dorsal hippocampus were randomly selected and assessed to determine if tracing criteria were met. Cells qualified for tracing if there was a clearly visible cell soma, a well impregnated apical dendrite and dendritic spines were visible. All spines were counted along all branches of the apical dendrite and dendritic spine density was determined by dividing the total number of spines by the total length of the apical dendrite tree.

Statistical Analysis

All data analysis was performed using Statistical Package for the Social Sciences (ver. 17 SPSS Inc., IL, USA). Maternal data was analysed using unpaired t tests. Water maze escape trials were analysed using three-way repeated measures ANOVA with Treatment, Sex, Age and Day as Factors. All other behavioural and histological data was analysed using three-way ANOVA with Treatment, Sex and Age as factors. Post hoc analysis with a Bonferroni correction was used where significant interactions between factors were observed. The observed power for analysis of stereology and dendrite morphology data was 0.88 and 0.77 respectively.

Results

Maternal data

We have previously reported maternal variables pertaining to the animals included in this study [45,46]. In brief, *ad libitum* consumption of this diet by pregnant dams results in a peak plasma ethanol concentration (PEC) of $\sim 0.03 \pm 0.01\%$ (30 mg/dl) approximately 30 minutes after offering fresh diet [45]. There were no significant differences in maternal diet consumption (28.3 \pm 0.8; 30.6 \pm 0.8 ml/day), total weight gain (97.5 \pm 6; 97.4 \pm 5 g), gestation length or litter size (10 \pm 1; 11 \pm 1) between dams fed an ethanol containing diet (EtOH) and dams fed an isocaloric control diet (Control) respectively [45,46].

Y Maze

Spatial memory performance was assessed in the Y maze task by analysing novel arm exploration (Figure 1). There were no significant effects of Treatment or Sex or interactions between factors for total arm entries, percentage of entries into the novel arm or percentage of time spent in the novel arm. However, a significant main effect of Age was found for total arm entries (F(1,80) = 34.44, P<0.001) and percentage time spent in the novel arm (F(1,80) = 6.53, P<0.05) with Aged animals making more total arm entries and spending a greater percentage of time within the novel arm than Adult animals.

Water Maze

The daily average latency to escape was analysed as a measure of learning in the water maze task. This data was analysed using three-way repeated measures ANOVA but has been graphically represented separated by age and sex (Figure 2). There was a significant overall effect of Day (F(1,80) = 91.41, P < 0.0001). There were no significant main effects of Treatment (F(1,80) = 0.24, P > 0.05), Age (F(1,80) = 0.01, P > 0.05) or Sex (F(1,80) = 0.13, P > 0.05) or significant interactions between these factors.

Spatial memory was assessed using the percentage time spent in the target quadrant and number of "platform crosses" during the probe trial (Table 1). There was no main effect of Treatment (F(1,80) = 0.18, P>0.05; F(1,80) = 0.33, P>0.05), Age (F(1,80) = 0.37, P>0.05; F(1,80) = 3.71, P>0.05) or Sex (F(1,80) = 2.26, P>0.05; F(1,80) = 2.17, P>0.05) for percentage time spent in the target quadrant or number of "platform crosses" respectively. No significant interactions were found between factors for "platform crosses" but a significant Sex*Age (F(1,80) = 3.15, P<0.05) interaction was found for percentage time in the target quadrant. However, a Bonferroni post test revealed no significant differences between groups.

Stereology

A significant main effect of Age was found for pyramidal cell number within the CA1 region (F(1,40) = 5.618, p < 0.05) where Aged animals had a greater estimated number of pyramidal cells than Adult animals (Figure 3a). A significant main effect of Sex was found for CA1 region volume (F(1,40) = 4.823, p < 0.05), where male animals had greater volume of the CA1 region than female animals (Figure 3c). No other main effects of Age, Sex or Treatment or interactions between factors were found for stereological measures within the CA1 region.

A significant main effect of Sex was found for CA3 volume (F(1,40) = 21.92, p < 0.01) and pyramidal cell number (F(1,40) = 21.62, p < 0.01) within this region with Male animals



Figure 1. Spatial working memory performance in a Y Maze task. n = 12-15 Aged rats and n = 10 Adult animals per sex, per treatment were included in the analysis for this task. Main effects determined by 3-way ANOVA are indicated by p values above each graph. a) Aged animals made significantly more arm entries overall than Adult animals. b) There were no significant effects of age, sex or prenatal treatment on % entries into the novel arm. c) Aged animals spent a greater % time in the novel arm than Adult animals. There were no significant effects of sex or prenatal treatment on % time spent in the novel arm.

doi:10.1371/journal.pone.0101482.g001

having a greater number of pyramidal cells and CA3 volume than Female animals (Figure 3b, d). However, Female animals were found to have greater cell density (F(1,40) = 4.96, p<0.05) than Male animals within the CA3 region. No other main effects or interactions were found for stereological measures within the CA3 region.

Apical Dendrite Morphology

A significant main effect of Age was found for apical dendrite length (F(1,40) = 5.98, p<0.05) with Aged animals having shorter apical dendrite length than Adult animals (Figure 4a). A main effect of Age was also found for the total number of branch points (F(1,40) = 12.41, p<0.01) for pyramidal cells in the CA1 region with Aged animals having less branching of pyramidal cell apical dendrites than Adult animals (5±1; 7±1 respectively). A significant interaction between Age, Sex and Treatment was also found for spine density (F(1,40) = 4.35, p = 0.049; Figure 4e). However, Bonferroni post-test analysis revealed no significant differences between groups. No other significant main effects or interactions were found for apical dendrite length, branch number, total spine number or spine density for pyramidal cell dendrites in the CA1 region.

No significant main effects or interactions were found for apical dendrite length, branch number or spine number for pyramidal cells within the CA3 region. However, a significant main effect of Treatment (F(1,40) = 7.53, p<0.05) and an Age*Sex*Treatment interaction (F(1,40) = 14.05, p=0.001) were found for spine density within this region (Figure 4f). Bonferroni post-test analysis showed EtOH Adult Male animals had a strong trend towards reduced spine density than Control Adult Male animals (F(1,20) = 6.07, p = 0.057) and that Ethanol exposed (EtOH) Aged Females had a significantly reduced spine density along the apical dendrite compared to their Control counterparts (F(1,20) = 11.65, p<0.05).

Discussion

The aim of this study was to investigate the long term consequences of low dose prenatal ethanol exposure on hippocampal-dependent spatial behaviour and morphology within the CA1 and CA3 regions of the hippocampus. Prenatal exposure to a liquid diet containing 6% (vol/vol) ethanol did not significantly impact spatial learning or memory performance in the water maze or Y maze tasks in either Adult or Aged rats of either sex. Similarly, our chronic low dose model of prenatal ethanol exposure did not induce alterations in pyramidal cell number or density in the CA1 or CA3 regions of the dorsal hippocampus. Interestingly, there was a subtle decrease in apical dendrite spine density for CA3 pyramidal cells of ethanol exposed animals that did not occur within the CA1 region. This may be indicative of a reduction in synaptic connectivity and activity within this region [45,46], that is not sufficient to induce overt deficits in hippocampal-dependent learning and memory performance.

The Y maze is often used to investigate spatial memory function in rodent models of acute and chronic stress [48,49]. Few studies however, have utilised the Y maze task in prenatal alcohol exposure models. The results from our study suggest that chronic low dose prenatal ethanol exposure does not affect spatial working memory in adult or aged rats. Interestingly, Dobson et al [55] utilised an adapted Y maze protocol as well as a dry land version of the Biel Maze task to assess spatial learning and memory in Guinea Pigs exposed to moderately high amounts of ethanol prenatally. They found deficits in performance in the dry land Biel Maze but, similar to our study, no deficits in performance in the Y maze task in their ethanol exposed animals. The authors conclude that the complexity of the dry land Biel Maze may be more sensitive to elucidating prenatal ethanol induced deficits in spatial learning and memory than the relatively simple two arm choice paradigm of the Y maze task [55].

In contrast to the infrequent utilisation of the Y maze task, many rodent models of FASD have employed variations of the Morris water maze paradigm to demonstrate deficits in spatial learning and memory function [28,30,31,56]. Differing from the chronic low dose consumption model utilised in our study, the majority of these studies model moderate to high dose exposure to ethanol and often employ a binge type exposure paradigm. Furthermore, dose response investigations of prenatal alcohol exposure have shown a dose dependent relationship between peak maternal blood ethanol concentrations and the severity of learning and memory deficits in offspring, often finding no significant



Figure 2. Spatial learning in the fixed platform water maze task. n = 12-15 Aged rats and n = 10 Adult animals per sex, per treatment were included in the analysis for this task. Data analysed using 3-way ANOVA but graphically separated for age and sex for visual clarity. There was a significant effect of day with all animals showing a reduction in time to escape over the four days. There were no significant effects of age, sex or prenatal treatment or interactions between these factors. doi:10.1371/journal.pone.0101482.g002

impairment at very low doses [37,57]. However, Savage and colleagues [38] demonstrated that prenatal exposure to a liquid diet containing 3% (vol/vol) ethanol resulting in similar peak maternal blood alcohol levels as our model (~30 mg/dl), was enough to reduce improvement in escape latency between trials in a moving platform Morris water maze task in adult male Sprague-Dawley rat offspring. Similarly, Sutherland and colleagues [58] demonstrate deficits in learning and memory performance in a moving platform Morris water maze paradigm after prenatal exposure to a 5% (vol/vol, 26% EDC) liquid diet in adult Sprague-Dawley rat offspring. It is interesting to note however, that both of these studies also show that in a fixed platform Morris

water maze task, similar to the one employed in our study, these offspring did not differ from control offspring in escape latency or probe trial performance [38,58]. This suggests that low dose prenatal ethanol exposure may not affect basic spatial learning and memory function but that more complex learning and memory processes may be impaired.

While many studies have shown regional cell loss in the hippocampus following prenatal alcohol exposure [40,59,60], we did not find any differences in pyramidal cell number or density in the CA1 or CA3 regions of the dorsal hippocampus in ethanol exposed offspring. However, other studies have shown no difference in cell number and density within the CA1 and CA3

Variable	Adult (11 months)		Aged (18 months)	
	Male	Female	Male	Female
Time in Target Quadrant (%):				
Control	32.5±2.4	30.2±2.3	29.7±1.9	32.4±1.9
EtOH	29.6±2.4	31.5±2.4	41.5±4.7	31.9±2.0
Platform Crosses:				
Control	3±0.5	3±0.4	3.3±0.4	2.2±0.4
EtOH	3.1±0.5	3.5±0.5	3.0±0.6	2.4±0.4

Table 1. Spatial memory performance in the water maze probe trial.

All data presented as mean±SEM.

doi:10.1371/journal.pone.0101482.t001



Figure 3. Stereological analysis of pyramidal neurons within the CA1 and CA3 regions of the dorsal hippocampus. Data from n = 6 brains per sex, age and prenatal treatment were included in the stereological analysis. a) Aged animals had a greater total number of pyramidal cells within the CA1 region than Adult animals. c) Male animals had a greater CA1 volume than Female animals. No significant differences were found for pyramidal cell density (e) within the CA1 region. b) Male animals had a greater estimated number of pyramidal neurons and a greater volume (d) of the CA3 region than Female animals. Female animals had a greater cell density within the CA3 region than Male animals. doi:10.1371/journal.pone.0101482.g003

regions in neonatal (PN10) and adult (PN112) rat offspring exposed to moderate to high levels of alcohol during the gestational period equivalent to the first two trimesters of human pregnancy (G1-22/23), the same exposure period used in our study [61,62]. Also similar to our findings, Tran and Kelly [61] show greater pyramidal cell numbers in male animals than female animals. However, the gender difference in the Tran and Kelly [61] study was consistent throughout the CA1, CA3 and Dentate Gyrus, while in our study the gender difference in pyramidal cell number was only significant in the CA3 region. Furthermore, mouse models using a similar exposure period have also shown a null effect of moderate prenatal alcohol exposure and chronic alcohol exposure on hippocampal-dependent behaviour or hippocampal morphometry [63,64].

Perhaps some of the most intriguing results from our study come from the spine density along pyramidal cell apical dendrites within the CA3 region. Previous research has demonstrated reductions in dendritic spine density as well as dendritic branching within the hippocampus, particularly within the CA1 region and the Dentate Gyrus [42,43,60]. We report a main effect of prenatal treatment for spine density within the CA3 region, suggesting an overall reduction in spine density for prenatal ethanol treated animals compared to control animals. This finding is lent some support by Tanaka and colleagues [41], who demonstrated a reduction in synaptic junctions within the CA3 region of near term rat pups (GD21) after maternal consumption of 5, 10 and 20% ethanol throughout gestation. However, we also report a significant interaction between prenatal treatment, age and sex for CA3 apical dendrite spine density in our animals. Previous studies have demonstrated recovery of prenatal alcohol induced synaptic deficits within the DG and CA1 region with age [42,43]. While there was a strong trend for a reduction in spine density in adult



Figure 4. Pyramidal cell apical Dendrite Morphology within the CA1 and CA3 regions of the dorsal hippocampus. Adult animals were found to have longer apical dendrites than Aged animals within the CA1 (a) but no such difference was found within the CA3 (b). There was no effect of age, sex or prenatal treatment on the total number of dendritic spines counted within the CA1 (c) or CA3 (d) region. e) A significant interaction between Age, Sex and Treatment was found for spine density within the CA1 region. However, Bonferroni post-test analysis revealed no significant differences between groups. f) Adult Male Control animals tended to have greater spine density than their EtOH exposed counterparts (indicated by line and #) and Aged Control Female animals had significantly greater density of dendritic spines along the apical dendrite than their EtOH exposed counterparts (indicated by line and *). doi:10.1371/journal.pone.0101482.g004

males following prenatal ethanol, this difference was not apparent in aged male animals. At face value, this might suggest that prenatal ethanol treated male offspring exhibit a similar recovery of spine density in aged animals compared to adult animals. However, this mitigation of the difference in spine density appears to be driven by a reduction in density in control animals while, spine density in ethanol male animals appears unchanged between age groups. Similarly, the apparent gain of spine density deficits in the CA3 region for aged prenatal ethanol treated females appears to be primarily driven by an increase in spine density in control female animals between the age groups with only a slight reduction in apical dendrite spine density in prenatal ethanol treated aged females.

Within the literature, administration of ethanol through a liquid diet is usually compared to a pair-fed control group to account for potential confounders associated with malnutrition [47]. Chow fed

controls have also been widely used within the literature to compare the effects of the diet or diet restriction (pair-fed), without alcohol, on the growth and development of offspring [38,65–67]. As the EtOH-exposed dams in the current study showed no sign of malnutrition, consumed the same amount of diet per day as the Control dams and both diets were nutritionally complete and approximately equivalent in caloric content, we felt that the inclusion of a specific pair-fed group was unnecessary. Equally, in the current study normal growth did not appear to be altered as the dams from both treatment groups showed similar weight gain patterns to those of a chow fed group (47 ± 3 g between E12–20) of the same rat strain, included in a previously study published by this laboratory [68].

Overall, within this study we did not find any deficits in spatial learning and memory ability or long term alterations in pyramidal cells within the CA1 and CA3 region of the dorsal hippocampus using a model of chronic low dose prenatal ethanol exposure. While the lack of deficits found within this study might seem somewhat reassuring to those who consume small amounts of alcohol during pregnancy, these findings should be taken in context. Our lab has previously reported subtle effects on growth and development in ethanol exposed offspring using this chronic low dose model [45]. Furthermore, there exists evidence of regional selectivity for ethanol-induced changes in the brain [69–71]. In line with this, we have recently shown that this model of chronic low dose prenatal ethanol exposure results in an increase in dendritic spine density and branching in the Basolateral (BLA), but not the medial (MeA) or central (CeA) nuclei of the amygdala in conjunction with a subtle phenotype for anxiety-like behaviour in these animals [46]. Collectively, these data suggest that

References

- Abel EL (1980) Fetal Alcohol Syndrome: Behavioral teratology. Psychological Bulletin 87: 29–50.
- Abel EL, Jacobson S, Sherwin BT (1983) In Utero Alcohol Exposure: Functional and Structural Brain Damage. Neurobehavioral Toxicology and Teratology 5: 363–366.
- Jones KL, Smith DW, Ulleland CN, Streissguth AP (1973) Pattern of Malformation in Offspring of Chronic Alcoholic Mothers. The Lancet 301: 1267–1271.
- Streissguth AP, Aase JM, Clarren SK, Randels SP, Ladue RA, et al. (1991) Fetal Alcohol Syndrome in Adolescents and Adults. Jama-Journal of the American Medical Association 265: 1961–1967.
- Abel EL (1995) An update on incidence of FAS: FAS is not an equal opportunity birth defect. Neurotoxicology and Teratology 17: 437–443.
- Kaminen-Ahola N, Ahola A, Maga M, Mallitt K, Fahey P, et al. (2010) Maternal Ethanol Consumption Alters the Epigenotype and the Phenotype of Offspring in a Mouse Model. PLoS Genet 6: e1000811.
- Streissguth AP, Bookstein FL, Sampson PD, Barr HM (1989) Neurobehavioral effects of prenatal alcohol: Part III. PLS analyses of neuropsychologic tests. Neurotoxicology and Teratology 11: 493–507.
- Streissguth AP, O'Malley K (2000) Neuropsychiatric implications and long-term consequences of fetal alcohol spectrum disorders. Seminars in Clinical Neuropsychiatry 5: 177–190.
- Riley EP, McGee CL (2005) Fetal alcohol spectrum disorders: An overview with emphasis on changes in brain and behavior. Experimental Biology and Medicine 230: 357–365.
- Famy C, Streissguth AP, Unis AS (1998) Mental Illness in Adults With Fetal Alcohol Syndrome or Fetal Alcohol Effects. American Journal of Psychiatry 155: 552–554.
- Barr HM, Bookstein FL, O'Malley KD, Connor PD, Huggins JE, et al. (2006) Binge Drinking During Pregnancy as a Predictor of Psychiatric Disorders on the Structured Clinical Interview for DSM-IV in Young Adult Offspring. American Journal of Psychiatry 163: 1061–1065.
- O'Connor MJ, Kasari C (2000) Prenatal Alcohol Exposure and Depressive Features in Children. Alcoholism: Clinical and Experimental Research 24: 1084–1092.
- Brown RT, Coles CD, Smith IE, Platzman KA, Silverstein J, et al. (1991) Effects of prenatal alcohol exposure at school age. II. Attention and behavior. Neurotoxicology and Teratology 13: 369–376.
- Carmichael Olson H, Streissguth AP, Sampson PD, Barr HM, Bookstein FL, et al. (1997) Association of Prenatal Alcohol Exposure with Behavioural and Learning Problems in Early Adolescence. J Am Acad Child Adolesc Psychiatry 36: 1187–1194.
- Rasmussen C, Becker M, McLennan J, Urichuk L, Andrew G (2011) An evaluation of social skills in children with and without prenatal alcohol exposure. Child Care Health and Development 37: 711–718.
- Hamilton DA, Kodituwakku P, Sutherland RJ, Savage DD (2003) Children with Fetal Alcohol Syndrome are impaired at place learning but not cued-navigation in a virtual Morris water task. Behavioural Brain Research 143: 85–94.
- Howell KK, Lynch ME, Platzman KA, Smith GH, Coles CD (2006) Prenatal alcohol exposure and ability, academic achievement, and school functioning in adolescence: A longitudinal follow-up. Journal of Pediatric Psychology 31: 116– 126.
- Kerns KA, Don A, Mateer CA, Streissguth AP (1997) Cognitive Deficits in Nonretarded Adults with Fetal Alcohol Syndrome. Journal of Learning Disabilities 30: 685–693.
- Sowell E, Lu LH, O'Hare D, McCourt ST, Mattson SN, et al. (2007) Functional magnetic resonance imaging of verbal learning in children with heavy prenatal alcohol exposure. Neuroreport 18: 635–639.
- Streissguth AP, Barr HM, Sampson PD (1990) Moderate Prenatal Alcohol Exposure: Effects on Child IQ and Learning Problems at Age 7^{1/2}. Alcoholism: Clinical and Experimental Research 14: 662–669.

consumption of small amounts of alcohol during pregnancy can lead to long term changes in offspring in some brain structures (BLA) while having no effect on others (hippocampus).

Acknowledgments

We would like to thank Suzy Alexander, Chelsea Stewart, Kim French, and Sorenta DeGasperi for their assistance in this study.

Author Contributions

Conceived and designed the experiments: CC TB NL KM. Performed the experiments: CC. Analyzed the data: CC TB. Contributed reagents/ materials/analysis tools: TB NL KM. Wrote the paper: CC TB NL KM.

- Willford JA, Richardson GA, Leech SL, Day N (2004) Verbal and Visuospatial Learning and Memory Function in Children with Moderate Prenatal Alcohol Exposure. Alcoholism: Clinical and Experimental Research 28: 497–507.
- Willoughby KA, Sheard ED, Nash K, Rovet J (2008) Effects of prenatal alcohol exposure on hippocampal volume, verbal learning and verbal and spatial recall in late childhood. Journal of the International Neuropsychological Society 14: 1022–1033.
- Streissguth AP, Sampson PD, Barr HM (1989) Neurobehavioral Dose-Response Effects of Prenatal Alcohol Exposure in Humans from Infancy to Adulthooda. Annals of the New York Academy of Sciences 562: 145–158.
- Mattson SN, Riley EP (1999) Implicit and explicit memory functioning in children with heavy prenatal alcohol exposure. Journal of the International Neuropsychological Society 5: 462–471.
- Osborne GL, Caul WF, Fernandez K (1980) Behavioral effects of prenatal ethanol exposure and differential early experience in rats. Pharmacology Biochemistry and Behavior 12: 393–401.
- Clausing P, Ferguson SA, Holson RR, Allen RR, Paule MG (1995) Prenatal ethanol exposure in rats: Long-lasting effects on learning. Neurotoxicology and Teratology 17: 545–552.
- Sanchez Vega MC, Chong S, Burne THJ (2013) Early gestational exposure to moderate concentrations of ethanol alters adult behaviour in C57BL/6J mice. Behavioural Brain Research 252: 326–333.
- Kelly SJ, Goodlett CR, Hulsether SA, West JR (1988) Impaired Spatial Navigation in Adult Female but not Adult Male-Rats Exposed to Alcohol During the Brain Growth Spurt. Behavioural Brain Research 27: 247–257.
- An L, Yang Z, Zhang T (2013) Imbalanced Synaptic Plasticity Induced Spatial Cognition in Male Offspring Rats Treated with Chronic Prenatal Ethanol Exposure. Alcoholism: Clinical and Experimental Research 37: 763–770.
- Iqbal U, Brien JF, Banjanin S, Andrews MH, Matthews SG, et al. (2005) Chronic prenatal ethanol exposure alters glucocorticoid signalling in the hippocampus of the postnatal guinea pig. Journal of Neuroendocrinology 17: 600–608.
- McAdam TD, Brien JF, Reynolds JN, Dringenberg HC (2008) Altered watermaze search behavior in adult guinea pigs following chronic prenatal ethanol exposure: Lack of mitigation by postnatal fluoxetine treatment. Behavioural Brain Research 191: 202–209.
- Schneider ML, Moore CF, Kraemer GW (2001) Moderate alcohol during pregnancy: Learning and behavior in adolescent rhesus monkeys. Alcoholism-Clinical and Experimental Research 25: 1383–1392.
- Nagahara AH, Handa RJ (1997) Fetal alcohol exposure produces delaydependent memory deficits in juvenile and adult rats. Alcoholism-Clinical and Experimental Research 21: 710–715.
- Gianoulakis C (1989) Rats exposed prenatally to alcohol exhibit impairments in spatial navigation test. Behavioural Brain Research 36: 217–228.
- Cronise K, Marino MD, Tran TD, Kelly SJ (2001) Critical Periods for the Effects of Alcohol Exposure on Learning in Rats. Behavioural Neuroscience 115: 138–145.
- Lochry EA, Riley EP (1980) Retention of passive-avoidance and T-maze escape in rats exposed to alcohol prenatally. Neurobehavioral Toxicology 2: 107–115.
- Tomlinson D, Wilce P, Bedi KS (1998) Spatial Learning Ability of Rats Following Differing Levels of Exposure to Alcohol During Early Postnatal Life. Physiology & Behavior 63: 205–211.
- Savage DD, Becher M, de la Torre AJ, Sutherland RJ (2002) Dose-dependent effects of prenatal ethanol exposure on synaptic plasticity and learning in mature offspring. Alcoholism-Clinical and Experimental Research 26: 1752–1758.
- Berman RE, Hannigan JH (2000) Effects of prenatal alcohol exposure on the hippocampus: Spatial behavior, electrophysiology, and neuroanatomy. Hippocampus 10: 94–110.
- Livy DJ, Miller EK, Maier SE, West JR (2003) Fetal alcohol exposure and temporal vulnerability: effects of binge-like alcohol exposure on the developing rat hippocampus. Neurotoxicology and Teratology 25: 447–458.

- Tanaka H, Nasu F, Inomata K (1991) Fetal Alcohol Effects: Decreased Synaptic Formatations in the Field CA3 of Fetal Hippocampus. International Journal of Developmental Neuroscience 9: 509–517.
- Hoff SF (1988) Synaptogenesis in the Hippocampal Dentate Gyrus: Effects of In Utero Ethanol Exposure. Brain Research Bulletin 21: 47–54.
- 43. Tarelo-Acuna L, Olvera-Cortes ME, Gonzalez-Burgos I (2000) Prenatal and postnatal exposure to ethanol induces changes in the shape of the dendritic spines from hippocampal CA1 pyramidal neurons of the rat. Neuroscience Letters 286: 13–16.
- 44. Sutherland RJ, McDonald RJ, Savage DD (1997) Prenatal Exposure to Moderate Levels of Ethanol Can Have Long-Lasting Effects on Hippocampal Synaptic Plasticity in Adult Offspring. Hippocampus 7: 232–238.
- 45. Probyn M, Zanini S, Ward L, Bertram J, Moritz K (2012) A rodent model of low- to moderate-dose ethanol consumption during pregnancy: patterns of ethanol consumption and effects on fetal and offspring growth. Reproduction, Fertility and Development 24: 859–870.
- Cullen CL, Burne THJ, Lavidis NA, Moritz KM (2013) Low Dose Prenatal Ethanol Exposure Induces Anxiety-Like Behaviour and Alters Dendritic Morphology in the Basolateral Amygdala of Rat Offspring, Plos One 8.
- Lieber CS, DeCarli LM (1982) The Feeding of Alcohol in Liquid Diets: Two Decades of Applications and 1982 Update. Alcoholism: Clinical and Experimental Research 6: 523-531.
- Conrad CD, Grote KA, Hobbs RJ, Ferayorni A (2003) Sex differences in spatial and non-spatial Y-maze performance after chronic stress. Neurobiology of Learning and Memory 79: 32–40.
- Dellu F, Mayo W, Cherkaoui J, Le Moal M, Simon H (1992) A two-trial memory task with automated recording: study in young and aged rats. Brain Research 588: 132–139.
- Morris R (1984) Developments of a water-maze procedure for studying spatial learning in the rat. Journal of Neuroscience Methods 11: 47–60.
- Morris RGM (1981) Spatial Location Does Not Require the Presence of Local Cues. Learning and Motivation 12: 239–260.
- 52. Paxinos G, Watson C (2006) The rat brain in stereotaxic coordinates: hard cover edition: Access Online via Elsevier.
- West MJ (1993) New Stereological Methods for Counting Neurons. Neurobiology of Aging 14: 275–285.
- 54. Gibb R, Kolb B (1998) A method for vibratome sectioning of Golgi–Cox stained whole rat brain. Journal of Neuroscience Methods 79: 1–4.
- Dobson CC, Mongillo DL, Poklewska-Koziell M, Winterborn A, Brien JF, et al. (2012) Sensitivity of modified Biel-maze task, compared with Y-maze task, to measure spatial learning and memory deficits of ethanol teratogenicity in the guinea pig. Behavioural Brain Research 233: 162–168.
- Richardson DP, Byrnes ML, Brien JF, Reynolds JN, Dringenberg HC (2002) Impaired acquisition in the watermaze and hippocampal long-term potentiation after chronic prenatal ethanol exposure in the guinea pig. European Journal of Neuroscience 16: 1593–1598.
- Reyes E, Wolfe J, Savage DD (1989) The effects of prenatal alcohol exposure on radial arm maze performance in adult rats. Physiology & Behavior 46: 45–48.

- Sutherland RJ, McDonald RJ, Savage DD (2000) Prenatal exposure to moderate levels of ethanol can have long-lasting effects on learning and memory in adult offspring. Psychobiology 28: 532.
- Miller MW (1995) Generation of Neurons in the Rat Dentate Gyrus and Hippocampus: Effects of Prenatal and Postnatal Treatment with Ethanol. Alcoholism: Clinical and Experimental Research 19: 1500–1509.
- Gonzalez-Burgos I, Alejandre-Gomez M, Olvera-Cortes ME, Perez-Vega MI, Evans S, et al. (2006) Prenatal-through-postnatal exposure to moderate levels of ethanol leads to damage on the hippocampal CA1 field of juvenile rats - A stereology and Golgi study. Neuroscience Research 56: 400–408.
- Tran TD, Kelly SJ (2003) Critical periods for ethanol-induced cell loss in the hippocampal formation. Neurotoxicology and Teratology 25: 519–528.
- Maier SE, West JR (2001) Regional differences in cell loss associated with bingelike alcohol exposure during the first two trimesters equivalent in the rat. Alcohol 23: 49–57.
- Mineur YS, Marican CCG, Larue-Achagiotis C, Sluyter F, Crusio WE (2007) Effects of Chronic Alcohol Consumption on Hippocampal Anatomy and Associated Behaviors in Three Inbred Strains of Mice. The Open Behavioral Science Journal 1: 5–12.
- 64. Sluyter F, Jamot L, Bertholet J, Crusio WE (2005) Prenatal exposure to alcohol does not affect radial maze learning and hippocampal mossy fiber sizes in three inbred strains of mouse. Behavioral and Brain Functions 1.
- 65. Brocardo PS, Bochme F, Patten A, Cox A, Gil-Mohapel J, et al. (2012) Anxietyand depression-like behaviors are accompanied by an increase in oxidative stress in a rat model of fetal alcohol spectrum disorders: Protective effects of voluntary physical exercise. Neuropharmacology 62: 1607–1618.
- Hoffman CE, Patyk IA, Weinberg J (2005) Prenatal ethanol exposure: Sex differences in anxiety and anxiolytic response to 5-HT_{1A} agonist. Pharmacology Biochemistry and Behavior 82: 549–558.
- Glavas MM, Ellis L, Yu WK, Weinberg J (2007) Effects of prenatal ethanol exposure on basal limbic-hypothalamic-pituitary-adrenal regulation: Role of corticosterone. Alcoholism-Clinical and Experimental Research 31: 1598–1610.
- Gray SP, Denton KM, Cullen-McEwen L, Bertram JF, Moritz KM (2010) Prenatal Eposure to Alcohol Reduces Nephron Number and Raises Blood Pressure in Progeny. Journal of the American Society of Nephrology 21: 1891– 1902.
- 69. Rice JP, Suggs LE, Lusk AV, Parker MO, Candelaria-Cook FT, et al. (2012) Effects of exposure to moderate levels of ethanol during prenatal brain development on dendritic length, branching, and spine density in the nucleus accumbens and dorsal striatum of adult rats. Alcohol 46: 577–584.
- Hamilton DA, Akers KG, Rice JP, Johnson TE, Candelaria-Cook FT, et al. (2010) Prenatal exposure to moderate levels of ethanol alters social behavior in adult rats: Relationship to structural plasticity and immediate early gene expression in frontal cortex. Behavioural Brain Research 207: 290–304.
- Ledig M, Ciesielski L, Simler S, Lorentz JG, Mandel P (1988) Effect of Pre- and Postnatal Alcohol Consumption on GABA Levels of Various Brain Regions in the Rat Offspring. Alcohol and Alcoholism 23: 63–67.