

Draft Genome Sequence of the First Isolate of Extensively Drug-Resistant *Mycobacterium tuberculosis* in Ireland

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Extensive drug resistance is an emerging threat to the control of tuberculosis (TB) worldwide, even in countries with low TB incidence. We report the draft whole-genome sequence of the first reported extensively drug-resistant TB (XDR-TB) strain isolated in Ireland (a low-incidence setting) and describe a number of single-nucleotide variations that correlate with its XDR phenotype.

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Multidrug resistance (MDR) in tuberculosis (TB) threatens the global management of the disease, which is already a leading cause of infectious mortality worldwide, with an estimated 450,000 MDR-TB cases reported in 2012 (1). Approximately 10% of MDR-TB cases (those resistant to rifampin and isoniazid) are further defined as extensively drug resistant (XDR)-TB, due to their resistance to second-line drugs, fluoroquinolones and injectable aminoglycosides (2). Long turnaround times (2 to 4 weeks) for phenotypic drug susceptibility testing (DST) (due to the fastidious nature of the organism) can hamper the appropriate treatment of XDR-TB by delaying access to antibiotic susceptibility data (3). Next-generation sequencing (NGS) can highlight resistance in a timely manner in order to effectively manage treatment and minimize further transmission of resistant strains (4–6).

The first Irish XDR-TB strain was isolated in the Irish Mycobacteria Reference Laboratory (IMRL) in 2005 (IEXDR1) (7, 8). First-line DST was completed within 3 weeks (found to be streptomycin, isoniazid, rifampin, ethambutol, and pyrazinamide resistant), second-line DST within 5 weeks (found to be amikacin, clarithromycin, ciprofloxacin, and rifabutin resistant, as well as capreomycin, clofazimine, and prothionamide susceptible), and the remainder within 14 weeks (found to be *para*-aminosalicylate sodium [PAS] resistant and ethionamide and cycloserine susceptible).

In 2014, NGS was performed to provide further molecular characterization of IEXDR1 (lineage 2 or Beijing strain). Genomic DNA was sequenced using an Illumina MiSeq. Paired-end reads were mapped to the *Mycobacterium tuberculosis* H37Rv reference genome (accession no. AL123456.3) using the Burrows-Wheeler Aligner (9). This yielded a mapped-read depth of 196-fold, covering 97.6% of the H37Rv genome. A consensus sequence was called using the SAMtools mpileup command (10). The IMAGE algorithm was employed to extend contigs and close gaps in the assembly, producing a final draft assembly of 4,340,174 bp, consisting of

109 contigs (11). Single-nucleotide polymorphism (SNP) analysis was performed using Geneious R7 (version 7.1.5; Biomatters); 1,492 SNPs were detected in the assembled genome with respect to the genome of H37Rv, of which 810 were nonsynonymous (depth of coverage, ≥ 20 -fold [average, 276]; variant frequency, $\geq 95\%$).

Nonsynonymous mutations were identified in genes Rv0667/*rpoB* (H526Y) and Rv1908c/*katG* (S315T). There is strong correlation between substitutions in *rpoB* (H526Y) and *katG* (S315T) and phenotypic resistance to rifampin and isoniazid, respectively (4, 12). High-confidence SNPs were also found for fluoroquinolone resistance in gene Rv0006 (*gyrA*) (D94A) and aminoglycoside resistance in MTB000019/*rrs* (a1401g) (12). This is consistent with the XDR phenotype of IEXDR1. Other high-confidence mutations found in IEXDR1 for ethambutol (Rv3795/*embB* [M306V]) and streptomycin (Rv0682/*rpsL* [K43R]) correlate with its drug resistance profile (13, 14). SNPs that may confer resistance to pyrazinamide (Rv2043c/*pncA* [G132C]) and PAS (Rv3764c/*thyA* [Q97R]) were also identified, although their specificities and sensitivities are not as well defined (http://www.broadinstitute.org/annotation/genome/mtb_drug_resistance.1/DirectedSequencingHome.html).

Previously described phylogenetically informative polymorphisms (Rv1908c/*katG* [R463L], Rv2629 [D64A], Rv3794/*embA* [C76C, TGC/TGT] and [Q38Q, CAA/CAG], Rv1630/*rpsA* [R212R, CGA/CGC], Rv3919c/*gidB* [E92D], and Rv0486/*mshA* [A187V]) confirm the presence of a Beijing strain (15).

In summary, using NGS, this isolate was confirmed to be XDR-TB in a considerably shorter turnaround time than that for conventional DST. This underlines the potential of NGS in the diagnostic laboratory, especially for MDR- and XDR-TB cases.

Nucleotide sequence accession number. This whole-genome sequencing project has been deposited in the European Nucleotide Archive under the accession no. [CCJS00000000](https://www.ebi.ac.uk/ena/submit/).

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REFERENCES

1. World Health Organization. 2013. Global tuberculosis report. World Health Organization, Geneva, Switzerland.
2. CDC. 2006. Revised definition of extensively drug-resistant tuberculosis. *MMWR Morb. Mortal. Wkly. Rep.* 55:1176.
3. Campbell PJ, Morlock GP, Sikes RD, Dalton TL, Metchock B, Starks AM, Hooks DP, Cowan LS, Plikaytis BB, Posey JE. 2011. Molecular detection of mutations associated with first- and second-line drug resistance compared with conventional drug susceptibility testing of *Mycobacterium tuberculosis*. *Antimicrob. Agents Chemother.* 55:2032–2041. <http://dx.doi.org/10.1128/AAC.01550-10>.
4. Köser CU, Bryant JM, Becq J, Török ME, Ellington MJ, Marti-Renom MA, Carmichael AJ, Parkhill J, Smith GP, Peacock SJ. 2013. Whole-genome sequencing for rapid susceptibility testing of *M. tuberculosis*. *N. Engl. J. Med.* 369:290–292. <http://dx.doi.org/10.1056/NEJMc1215305>.
5. Köser CU, Ellington MJ, Cartwright EJ, Gillespie SH, Brown NM, Farrington M, Holden MT, Dougan G, Bentley SD, Parkhill J, Peacock SJ. 2012. Routine use of microbial whole genome sequencing in diagnostic and public health microbiology. *PLoS Pathog.* 8:e1002824. <http://dx.doi.org/10.1371/journal.ppat.1002824>.
6. Wyres K, Conway T, Garg S, Queiroz C, Reumann M, Holt K, Rusu L. 2014. WGS analysis and interpretation in clinical and public health microbiology laboratories: what are the requirements and how do existing tools compare? *Pathogens* 3:437–458. <http://dx.doi.org/10.3390/pathogens3020437>.
7. McLaughlin AM, O'Donnell RA, Gibbons N, Scully M, O'Flanagan D, Keane J. 2007. Extensively drug-resistant tuberculosis (XDR-TB)—a potential threat in Ireland. *Open Respir. Med. J.* 1:7–9. <http://dx.doi.org/10.2174/1874306400701010007>.
8. Kennedy B, Lyons O, McLoughlin AM, Gibbons N, O'Flanagan D, Keane J. 2008. Extensively drug-resistant tuberculosis: first report of a case in Ireland. *Euro Surveill.* 13:pii = 18935. <http://www.eurosurveillance.org/ViewArticle.aspx?ArticleId=18935>.
9. Li H, Durbin R. 2009. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics* 25:1754–1760. <http://dx.doi.org/10.1093/bioinformatics/btp324>.
10. Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, 1000 Genome Project Data Processing Subgroup. 2009. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 25:2078–2079. <http://dx.doi.org/10.1093/bioinformatics/btp352>.
11. Swain MT, Tsai IJ, Assefa SA, Newbold C, Berriman M, Otto TD. 2012. A post-assembly genome-improvement toolkit (PAGIT) to obtain annotated genomes from contigs. *Nat. Protoc.* 7:1260–1284. <http://dx.doi.org/10.1038/nprot.2012.068>.
12. Rodwell TC, Valafar F, Douglas J, Qian L, Garfein RS, Chawla A, Torres J, Zadorozhny V, Kim MS, Hoshide M, Catanzaro D, Jackson L, Lin G, Desmond E, Rodrigues C, Eisenach K, Victor TC, Ismail N, Crudu V, Gler MT, Catanzaro A. 2014. Predicting extensively drug-resistant *Mycobacterium tuberculosis* phenotypes with genetic mutations. *J. Clin. Microbiol.* 52:781–789. <http://dx.doi.org/10.1128/JCM.02701-13>.
13. Sreevatsan S, Stockbauer KE, Pan X, Kreiswirth BN, Moghazeh SL, Jacobs WR, Jr, Telenti A, Musser JM. 1997. Ethambutol resistance in *Mycobacterium tuberculosis*: critical role of *embB* mutations. *Antimicrob. Agents Chemother.* 41:1677–1681.
14. Nair J, Rouse DA, Bai GH, Morris SL. 1993. The *rpsL* gene and streptomycin resistance in single and multiple drug-resistant strains of *Mycobacterium tuberculosis*. *Mol. Microbiol.* 10:521–527. <http://dx.doi.org/10.1111/j.1365-2958.1993.tb00924.x>.
15. Feuerriegel S, Köser CU, Niemann S. 2014. Phylogenetic polymorphisms in antibiotic resistance genes of the *Mycobacterium tuberculosis* complex. *J. Antimicrob. Chemother.* 69:1205–1210. <http://dx.doi.org/10.1093/jac/dkt535>.