airway oxidative stress and inflammation in asthmatic children.

# Acute Effects of Air Pollution on Pulmonary Function, Airway Inflammation, and Oxidative Stress in Asthmatic Children

Ling Liu,<sup>1</sup> Raymond Poon,<sup>1</sup> Li Chen,<sup>1</sup> Anna-Maria Frescura,<sup>1</sup> Paolo Montuschi,<sup>2</sup> Giovanni Ciabattoni,<sup>2</sup> Amanda Wheeler,<sup>1</sup> and Robert Dales<sup>1</sup>

<sup>1</sup>Healthy Environments and Consumer Safety Branch, Health Canada, Ottawa, Ontario, Canada; <sup>2</sup>Department of Pharmacology, Faculty of Medicine, Catholic University of the Sacred Heart, Rome, Italy

BACKGROUND: Air pollution is associated with respiratory symptoms, lung function decrements, and hospitalizations. However, there is little information about the influence of air pollution on lung injury.

OBJECTIVE: In this study we investigated acute effects of air pollution on pulmonary function and

METHODS: We studied 182 children with asthma, 9–14 years of age, for 4 weeks. Daily ambient concentrations of sulfur dioxide, nitrogen dioxide, ozone, and particulate matter  $\leq 2.5~\mu m$  in aerodynamic diameter (PM $_{2.5}$ ) were monitored from two stations. Once a week we measured spirometry and fractional exhaled nitric oxide (Fe $_{NO}$ ), and determined thiobarbituric acid reactive substances (TBARS) and 8-isoprostane—two oxidative stress markers—and interleukin-6 (IL-6) in breath condensate. We tested associations using mixed-effects regression models, adjusting for confounding variables.

RESULTS: Interquartile-range increases in 3-day average SO $_2$  (5.4 ppb), NO $_2$  (6.8 ppb), and PM $_{2.5}$  (5.4 µg/m³) were associated with decreases in forced expiratory flow between 25% and 75% of forced vital capacity, with changes being -3.1% [95% confidence interval (CI), -5.8 to -0.3], -2.8% (95% CI, -4.8 to -0.8), and -3.0% (95% CI, -4.7 to -1.2), respectively. SO $_2$ , NO $_2$ , and PM $_{2.5}$  were associated with increases in TBARS, with changes being 36.2% (95% CI, 15.7 to 57.2), 21.8% (95% CI, 8.2 to 36.0), and 24.8% (95% CI, 10.8 to 39.4), respectively. Risk estimates appear to be larger in children not taking corticosteroids than in children taking corticosteroids. O $_3$  (5.3 ppb) was not associated with health end points. Fe $_{\rm NO}$ , 8-isoprostane, and IL-6 were not associated with air pollutants.

CONCLUSION: Air pollution may increase airway oxidative stress and decrease small airway function of asthmatic children. Inhaled corticosteroids may reduce oxidative stress and improve airway function.

KEY WORDS: air pollution, asthma, children, exhaled breath condensate, inflammation, oxidative stress, pulmonary function. *Environ Health Perspect* 117:668–674 (2009). doi:10.1289/ehp11813 available via *http://dx.doi.org/* [Online 28 November 2008]

Asthma is one of the most prevalent chronic health conditions in Canadian children. The 1998–1999 National Population Health Survey found that the prevalence of physician-diagnosed asthma is 8.4% among the whole population and 10.7% among children and teenagers, corresponding to approximately 845,000 children (Canadian Institute for Health Information et al. 2001). Asthma compromises the quality of life for patients and their families and imposes a burden on the nation's health care expenditure. In 1990, the economic costs of asthma for Canada were estimated to fall between \$504 and \$648 million (Krahn et al. 1996).

Epidemiologic studies have shown that air pollution is associated with asthma-related increases in hospital admissions (Burnett et al. 1999; Wong et al. 2001), emergency department visits (Stieb et al. 2000; Sunyer et al. 1997; Villeneuve et al. 2007), and respiratory symptoms (Brauer et al. 2002; Vichit-Vadakan et al. 2001), and decreases in pulmonary function (Delfino et al. 2004; Gielen et al. 1997). However, there is a dearth of information about whether these associations can be attributed to lung tissue injury in an asthmatic population, partly because of the lack of appropriate non-invasive biomarkers

indicating lung inflammation and tissue injury. In recent years, some researchers have used fractional exhaled nitric oxide (Fe<sub>NO</sub>) to investigate the relationship between exposure to air pollutants and airway inflammation (Delfino et al. 2006; Mar et al. 2005).

Measurements of mediators in exhaled breath condensate have been used to better understand the pathological mechanisms of airway diseases such as asthma, chronic obstructive pulmonary disease, and cystic fibrosis (Horvath et al. 2005; Montuschi 2005). Recently, analysis of mediators in condensed breath has been extended to include lung diseases caused by occupational exposure (Lehtonen et al. 2007; Pelclová et al. 2007). Asthma is a chronic airway inflammatory disease involving a variety of inflammatory cell types and proinflammatory mediators. Reactive oxygen species are also thought to play an active role in inducing airway inflammation and hyperresponsivness (Hulsmann et al. 2007). Reactive oxygen species result from an increased burden of exogenous oxidants, endogenous lipid peroxidation, as well as from inflammatory cells (Halliwell and Gutteridge 1989). The level of reactive oxygen species in the airways can be determined by assaying concentrations of oxidative stress markers in breath condensate (Horvath et al. 2005).

We designed this longitudinal study to test the hypothesis that an acute increase in ambient air pollutants is associated with increased levels of markers of oxidative stress and inflammation in breath condensate of children with asthma, and that changes of breath markers are consistent with changes in pulmonary function and Fe<sub>NO</sub>.

# **Materials and Methods**

Study subjects. The study protocol was approved by the Research Ethics Board of Health Canada. The parent of each child gave written informed consent before the child's participation in this study.

The study was conducted between October and December of 2005 in Windsor, Ontario, Canada. Asthmatic children, 9 to 14 years of age, from nonsmoking households, were identified from a questionnaire survey of grades one to eight of elementary schoolchildren whose parents were asked whether or not the child had doctor-diagnosed asthma. The parents who gave a positive response were telephoned to confirm the child's current status of doctor-diagnosed asthma and whether there was any smoking of tobacco products in the household. One hundred eighty two subjects agreed to participate in this longitudinal study. Information on family income and family asthma history was extracted from the survey questionnaire. For logistical purposes, children were studied in two test periods, either between 11 October and 11 November (95 children), or between 14 November and 9 December 2005 (87 children). We did not advise parents to alter their children's daily activity and the use of asthma medications.

Ambient air pollution monitoring. Hourly measures of particulate matter  $\leq 2.5 \mu m$  in

Address correspondence to L. Liu, Health Canada. A.L. 4903B, 269 Laurier Ave., Ottawa, Ontario, Canada K1A 0K9. Telephone: (613) 954-4457. Fax: (613) 948-8482. E-mail: ling\_liu@hc-sc.gc.ca

Supplemental Material is available online at http:// www.ehponline.org/members/2008/11813/suppl.pdf We gratefully acknowledge S. De Pauw of Health Canada for coordinating the study, B. Nadeau and

Canada for coordinating the study, B. Nadeau and K.L. Ku for running laboratory assays, M. Smith-Doiron for preparing the map, and D. Stieb and P. Villeneuve for reviewing the manuscript.

This study was funded by the Canada-the United States Border Air Quality Strategy.

The authors declare they have no competing financial interests.

Received 17 June 2008; accepted 28 November 2008.

aerodynamic diameter (PM<sub>2.5</sub>), sulfur dioxide, nitrogen dioxide, and ozone were available from the National Air Pollution Surveillance network of Environment Canada (http://www. etc-cte.ec.gc.ca/NapsData). There were two monitoring stations representing Windsor, located upwind from the prevailing winds. Of 182 participating children, 180 of them lived within a 10-km radius from the air monitoring sites, and one lived 12 km and another 17 km from the air sampling sites (Figure 1). Values from these two monitors were averaged. SO2 was measured using ultraviolet fluorescence and the electrical conductivity from changes in chemical composition of a bromine solution, NO<sub>2</sub> measured using chemiluminescence, O<sub>3</sub> measured using ultraviolet absorbance, and PM<sub>2.5</sub> measured using tapered element oscillating microbalance monitor. Pollutant concentrations were averaged over a 24-hr period.

Measurement of spirometry and  $Fe_{NO}$ . Once a week, registered respiratory technologists visited the children's schools, and conducted tests of spirometry and  $Fe_{NO}$  and collected exhaled breath condensate on study participants. These tests were run on the same day of the week for each child.

Spirometry was performed according to American Thoracic Society criteria (American Thoracic Society 1995). KoKo Spirometers (Ferraris CardioRespiratory; Pulmonary Data Services, Inc. Louiseville, CO, USA) were calibrated daily and results adjusted for temperature and barometric pressure. A maximum of eight forced vital capacity (FVC) maneuvers were carried out in an attempt to achieve three acceptable flow-volume loops and two being within 0.20 L for FVC and forced expiratory volume in one second (FEV<sub>1</sub>), and within 0.20 L/sec for the mean forced expiratory flow between 25% and 75% of the FVC (FEF<sub>25-75%</sub>). The value assigned to the child was the largest acceptable value within 200 mL of a second value.

Single-breath on-line measures of  $Fe_{NO}$ were carried out according to American Thoracic Society and European Respiratory Society Guidelines (American Thoracic Society 2005) using an Eco Physics CLD AL MED chemiluminescence analyzer (Eco Medics AG, Duernten, Switzerland). Before performing a slow vital capacity maneuver over at least 6 sec at 0.05 L/sec, subjects took three tidal volume breaths through a DENOX 88 (Eco Medics AG, Duernten, Switzerland) which contains an NO<sub>x</sub> absorber cartridge to scrub the ambient air to a value of between 0 and 1.5 ppb. The test was repeated a maximum of eight times in an attempt to obtain at least two acceptable plateau Fe<sub>NO</sub> values within 10%. The value assigned to the subject was the mean of the two values. We calculated Fe<sub>NO</sub> statistics using spiroWare 88 software (Eco Medics AG, Duernten, Switzerland).

Collection of exhaled breath condensate, and measurement of amylase, oxidative stress, and inflammation. Exhaled breath condensate was collected using an RTube (Respiratory Research Inc., Charlottesville, VA, USA). Participants sat and breathed at tidal volumes orally into a mouthpiece attached to a cold condenser, for 10 min. Approximately 1 mL of breath condensate was collected. The condensed breath was then transferred into a microtube and stored at -20°C and then -80°C until use.

To determine saliva contamination, we measured  $\alpha$ -amylase activity in breath condensate using an assay kit (Pointe Scientific, MI, USA), which follows the increase in absorbance at 405 nm as the artificial substrate 2-chloro-p-nitrophenyl- $\alpha$ -D-maltotrioside is hydrolyzed.

Thiobarbituric acid reactive substances (TBARS) were determined in duplicate by a fluorescence method adapted from Yagi (1982). We used malonaldehyde bis-(dimethyl acetal) (Aldrich Chemical Co., Milwaukee, WI, USA) to construct a standard curve. To reduce the background interference with the very low levels of TBARS present in exhaled breath condensates, all glassware was washed in soap-free detergent followed by rinsing with distilled water and ethanol before use. We used butanol (OmniSolv, 99.95%; EM Science, Darmstadt, Germany) for extraction. The *n*-butanol extract was transferred to a Whatman glass bottom microplate (Whatman, NJ, USA), previously washed with ethanol, and the fluorescence measured at 530 nm/590 nm, excitation/emission, with a Cytofluor 2350 reader (Millipore, Billerica, MA, USA). The detection limit was 0.02 µM (three times the standard deviation above the blank). The within-run coefficient of variation was 8%.

A colorimetric enzyme immunoassay kit (Cayman Chemical, Ann Arbor, MI, USA) was used to measure 8-isoprostane concentrations in breath condensate. The assays were done in triplicate. A modified standard curve for 8-isoprostane was established for concentrations between 0.975 and 62.5 pg/mL. We were able to detect 8-isoprostane at a concentration as low as 0.975 pg/mL (three times the standard deviation above the blank). The within-run coefficient of variation was 5%. We also measured 8-isoprostane on 8 random samples using radioimmunoassay, a procedure previously described by Montuschi et al. (2003), and compared the results from both methods [see Supplemental Material, Table 1 (available online at http://www.ehponline.org/ members/2008/11813/suppl.pdf)]. Pearson correlation coefficient for results from two methods was 0.744 (p = 0.055), with a mean positive bias of approximately 35% in favor of the radioimmunoassay. Such a difference may be attributed to variations in the avidity and specificity of the antibodies used and the inherent variation during extrapolation from calibration curves at very low levels that were typical of exhaled breath condensates.

We determined the level of interleukin-6 (IL-6) using an ELISA kit from R&D Systems (Minneapolis, MN, USA). We conducted the ELISA assays in duplicate by following manufacturer's instructions accompanying the kit. The detection limit was 0.48 pg/mL.

Statistical analysis. We analyzed the correlations among environmental factors, daily use of inhaled corticosteroids (ICS) or short-acting

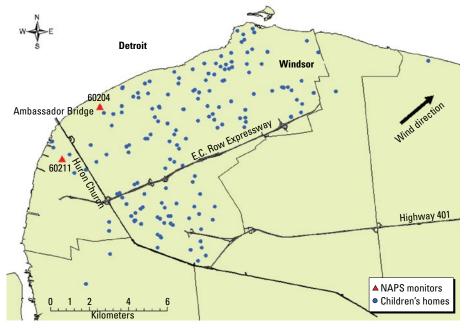


Figure 1. Map of Windsor, Ontario, Canada.

β-agonist (SABA), and testing period, using Spearman rank order correlation, because some variables were not normally distributed. We used mixed-effects regression models to account for repeated measures and to analyze the associations between pollutants and health end points, assuming random slope and random intercepts, and accounting for intraindividual correlation using S-PLUS (version 6.1; Insightful Corporation, Seattle, WA, USA). Data for Fe $_{NO}$ , TBARS, and 8-isoprostane were log-transformed to fit a normal distribution. Because > 99% of the samples gave a value > 0 for TBARS and 8-isoprostane, and most of them were above the detection limit,

data points below the detection limit were not given special treatment. For exposure variables, we tested models for multiday moving averages to examine potentially delayed response. Models were adjusted for time-dependent covariates that were statistically associated with either the exposure or response. We adjusted for testing period (as an indicator variable) to reduce the impact of seasonality. We also included product terms for the interaction of pollutant concentrations with sex and with the use of ICS to test potential modifying effects of sex and ICS. Since subjects served as their own control, we did not adjust for factors which remained constant during the study

Table 1. Selected characteristics of 182 asthmatic children from Windsor, Ontario, Canada.

Characteristic	Value
Age (years) [median (range)]	11 (9–14)
Sex [no. (%)]	
Female	68 (37.4)
Male	114 (62.6)
Race [no. (%)]	
Caucasian	160 (87.9)
Other	22 (12.1)
Parental asthma [no. (%)]	, ,
No No	11 (6.0)
Yes	70 (38.5)
Unknown	101 (55.5)
Self-reported medication use <sup>a</sup> [no. (%)]	(
ICS	68 (37.4)
SABA	64 (35.2)
Other asthma medications	33 (18.1)
None	77 (42.3)
FEV <sub>1</sub> (L) [median (5th– 95th percentile)]	2.4 (1.7–3.3)
Percent predicted FEV <sub>1</sub> (%) [median (5th– 95th percentile)] <sup>b</sup>	90.5 (68.6–107.1)
FEF <sub>25–75%</sub> (L/sec) [median (5th– 95th percentile)]	2.5 (1.3–3.9)
Percent predicted FEF <sub>25–75%</sub> (%) [median (5th– 95th percentile)] <sup>b</sup>	78.2 (45.3–118.0)
Fe <sub>NO</sub> (ppb) [median (5th– 95th percentile)]	19.4 (4.5–81.6)
TBARS (µM) [median (5th—95th percentile)]	0.26 (0.03–1.53)
8-Isoprostane (pg/mL) [median (5th– 95th percentile)]	1.43 (0.30–4.20)

<sup>a</sup>More than one medication could be used. <sup>b</sup>Predicted normal values were calculated based on a given height, age, sex, and race/ethnicity (Hankinson et al. 1999).

 Table 2. Values for selected environmental conditions during study period.

	Median [interquartile range (5th-95th percentile)]					
Environmental variable	1-day average	2-day average	3-day average			
SO <sub>2</sub> (ppb)	4.5 [6.5 (0.5 to 15.5)]	5.0 [5.6 (1.0 to 13.0)]	5.6 [5.4 (1.3 to 13.8)]			
NO <sub>2</sub> (ppb)	19.8 [9.8 (9.5 to 29.5)]	18.3 [9.0 (11.3 to 28.0)]	18.3 [6.8 (12.3 to 27.0)]			
O <sub>3</sub> (ppb)	13.0 [9.0 (6.5 to 26.5)]	14.1 [6.4 (6.8 to 23.3)]	14.0 [5.3 (7.5 to 21.0)]			
PM <sub>2.5</sub> (μg/m <sup>3</sup> )	6.5 [6.0 (2.0 to 19.0)]	6.6 [6.4 (2.0 to 15.0)]	6.9 [5.4 (2.7 to 14.3)]			
Temperature (°C)	6.6 [13.0 (-7.7 to 15.4)]	7.4 [12.0 (-5.5 to 14.7)]	7.4 [11.2 (-5.9 to 14.5)]			
Relative humidity (%)	67.4 [11.6 (53.3 to 87.9)]	65.6 [9.7 (54.4 to 75.6)]	65.7 [7.3 (54.9 to 74.2)]			

Air quality was monitored between 8 October and 9 December 2005, 62 sampling days.

Table 3. Spearman rank order correlation coefficients of variables.

Variable	Temperature	Relative humidity	SO <sub>2</sub>	NO <sub>2</sub>	03	PM <sub>2.5</sub>	SABA	ICS
Testing period Temperature Relative humidity SO <sub>2</sub> NO <sub>2</sub> O <sub>3</sub> PM <sub>2.5</sub> SABA	-0.73* /	0.01 0.18*	0.35* -0.17* -0.04	-0.30* 0.00 0.03 0.18*	0.06 0.18* -0.51* -0.02 -0.51*	-0.07 0.10* 0.20* 0.56* 0.71* -0.41*	0.10* -0.10* 0.01 0.04 -0.04 0.00 -0.05	-0.07 -0.02 -0.02 0.05 0.07 -0.10* 0.07 0.27*

<sup>\*</sup>p < 0.05.

period such as age, race, family income, and family asthma history. Because health testing was kept on the same day of week for each child, we did not adjust for the effect of day-of-week in the models.

When interactions were detected between pollutants and sex or the use of ICS, the participants were stratified for further analyses. We also ran two-pollutant models to adjust for potential confounding effects of co-pollutants. Results were expressed as percent change in FEV<sub>1</sub>, FEF<sub>25–75%</sub>, Fe<sub>NO</sub>, TBARS and 8-isoprostane per increase in interquartile-range concentration of a pollutant. A two-tailed value of  $p \le 0.05$  was considered statistically significant.

## Results

A total of 182 asthmatic children participated in the study, for a total of 691 spirometry test results, 672 Fe<sub>NO</sub> test results, and 616 breath samples. Seven children had 5 measurement days, 142 had 4 days, 27 had 3 days, and 6 had 2 days. In total, there were 39 measurement days during the study period, and no missing day for air monitoring to match health testing days. The characteristics of this cohort are presented in Table 1. We measured the concentration of amylase in 25% of breath samples to determine saliva contamination. The median amylase concentration was 0.60 unit/L, with the 95th percentile being 1.46 unit/L. None of the samples had amylase concentrations near the reported salivary amylase concentration of 1,250 unit/L (Brand et al. 2006). We estimated that saliva contamination was likely < 0.1%, and therefore interference of breath analysis by saliva was negligible. In 616 breath samples, only 2 samples had detectable IL-6 levels. We therefore did not conduct further analysis on IL-6.

The environmental conditions and daily concentrations of air pollutants measured during the study period are presented in Table 2. Because Spearman rank order correlation analyses showed that daily temperature, relative humidity and testing period were significantly correlated with some of the pollutants (Table 3), we included these variables in models to adjust for any confounding/modifying effects. We also included daily use of SABA and ICS to adjust for their potential confounding effects. Because pollutants were correlated with each other (Table 3), we also ran two-pollutant models to adjust for confounding.

Results from single-pollutant regression models. Table 4 presents percentage changes of health end points in relation to an increase in interquartile range concentrations of pollutants in a single-pollutant model which also included interaction terms of ICS and sex with pollutants. FEV<sub>1</sub> and FEF<sub>25–75%</sub> exhibited a consistent trend of negative associations with all four pollutants, with FEF<sub>25–75%</sub>

having a statistically significant association with NO<sub>2</sub> and PM<sub>2.5</sub>. The significant associations for FEF<sub>25-75%</sub> persisted for at least 3 days. Fe<sub>NO</sub> had a trend of positive associations with SO<sub>2</sub>, NO<sub>2</sub>, and PM<sub>2.5</sub>, but the associations were not statistically significant. Fe<sub>NO</sub> had a statistically significant negative association with O<sub>3</sub>. TBARS was significantly positively associated with SO<sub>2</sub>, NO<sub>2</sub>, and PM<sub>2.5</sub>, but not with O<sub>3</sub>. The associations for TBARS persisted for at least 3 days. 8-Isoprostane was significantly positively associated with same-day SO<sub>2</sub>, but not associated with other pollutants.

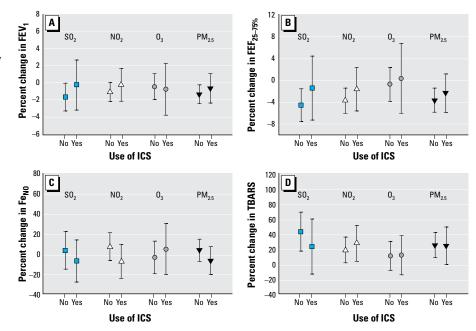
Modifying effects by the use of ICS and sex. Product terms between pollutants and the use of ICS showed significant interactions for some of the health end points. For example, the product term for SO<sub>2</sub> and ICS was positively associated with TBARS (p = 0.04), but negatively associated with FEV<sub>1</sub> (p < 0.01),  $FEV_{25-75\%}$  (p = 0.02), and 8-isoprostane (p = 0.01), indicating that the use of ICS was a significant effect modifier. The participants were then stratified by the use of ICS during the study period to adjust for modifying effects of use of ICS (Figure 2). Children who did not use ICS appeared to have larger risk estimates for pollutants than did those who used ICS. Among children not taking ICS, risk estimates for pollutants were statistically significant with respect to FEV<sub>1</sub>, FEF<sub>25-75%</sub>, and TBARS. Risk estimates for Fe<sub>NO</sub> and 8-isoprostane were not very different between the two groups of children (data not shown).

Product terms for pollutants and sex showed no significant interaction (p > 0.05), suggesting that sex was not a significant modifier for the associations between air pollutants

and health end points. We therefore did not run further analysis on boys and girls separately.

Results from two-pollutant models. Because we found consistent associations with pollutants for FEV<sub>1</sub>, FEF<sub>25-75%</sub>, Fe<sub>NO</sub>, and TBARS, we ran two-pollutant models on these end points to adjust for confounding effects of co-pollutants (Figure 3). For FEV<sub>1</sub>

and  $FEF_{25-75\%}$ , although two-pollutant models did not result in marked changes in risk estimates for  $PM_{2.5}$  and  $O_3$ , including  $PM_{2.5}$  in models did somewhat change risk estimates for  $SO_2$  and  $NO_2$ , suggesting that the effects of  $SO_2$  and  $NO_2$  on pulmonary function were influenced by  $PM_{2.5}$ . For  $Fe_{NO}$  and TBARS, including two pollutants in models did not markedly change the regression results,



**Figure 2.** Percent changes and 95% confidence intervals in  $FEV_1$  (A),  $FEF_{25-75\%}(B)$ ,  $Fe_{NO}$  (C), and breath TBARS (D) of asthmatic children taking or not taking ICS during the study period, in association with an interquartile-range increase in a pollutant. Sixty-eight asthmatic children took (Yes), and 114 children did not take (No) ICS. Pollutant concentrations were averaged over 3 days including the same day and 2 days before the clinical testing was done. Models were adjusted for testing period, daily temperature, relative humidity, and use of SABA.

Table 4. Percent change (95% confidence interval) in health end points of 182 asthmatic children, in association with an interquartile-range increase of a pollutant, in single-pollutant models with various lag times.

Air pollutant	FEV <sub>1</sub>	FEF <sub>25-75%</sub>	Fe <sub>NO</sub>	TBARS	8-Isoprostane
SO <sub>2</sub>					
Same day Lag 1 day 2-day average 3-day average	-0.3 (-1.3 to 0.7) 0.1 (-1.1 to 1.2) -0.1 (-1.4 to 1.3) -0.3 (-1.9 to 1.4)	-1.0 (-3.1 to 1.3) -0.9 (-3.1 to 1.4) -1.8 (-4.7 to 1.1) -2.3 (-5.5 to 1.0)	5.8 (-5.0 to 17.8) 1.6 (-9.3 to 13.9) 5.9 (-9.3 to 23.7) 1.7 (-14.1 to 20.3)	17.4 (0.3 to 37.4)* 14.1 (-1.8 to 32.5) 35.1 (9.5 to 66.8)* 61.8 (24.9 to 109.5)*	14.1 (2.5 to 26.9)* -2.4 (-11.7 to 7.9) 9.4 (-5.2 to 26.2) -0.3 (-16.4 to 18.9)
NO <sub>2</sub>	0.0 / 4.0 / 0.0	0.4/.4004)*	0.0 ( 0.0 ( 0.0 0)	04.0/4.0 . 44.0)*	0.0 / 40.7 / 40.4)
Same day Lag 1 day 2-day average 3-day average	-0.6 (-1.6 to 0.3) -0.3 (-1.1 to 0.6) -0.6 (-1.6 to 0.4) -0.8 (-1.9 to 0.3)	-2.4 (-4.3 to -0.4)* -1.4 (-3.1 to 0.3) -2.4 (-4.3 to -0.3)* -2.8 (-5.0 to -0.5)*	8.2 (-2.9 to 20.6) 3.7 (-6.1 to 14.6) 7.4 (-4.6 to 21.0) 0.5 (-12.1 to 14.9)	21.2 (1.9 to 44.2)* 10.2 (–5.6 to 28.7) 20.3 (–0.4 to 45.3) 32.9 (7.2 to 64.6)*	0.0 (-10.7 to 12.1) -5.6 (-14.8 to 4.5) -4.7 (-1.1 to 8.1) -4.0 (-16.3 to 10.2)
$0_{3}$					
Same day Lag 1 day 2-day average 3-day average	-0.4 (-1.6 to 0.7) -0.2 (-1.1 to 0.7) -0.4 (-1.6 to 0.9) -0.5 (-2.0 to 1.0)	0.2 (-2.1 to 2.5) 0.1 (-1.7 to 1.9) 0.4 (-2.2 to 3.0) -0.4 (-3.5 to 2.7)	-12.2 (-22.3 to -0.8)* -8.3 (-16.0 to 0.2) -16.0 (-26.4 to -4.1)* -1.6 (-16.7 to 16.1)	5.0 (-13.2 to 27.0) 7.3 (-6.7 to 23.4) 14.4 (-8.5 to 43.0) 8.8 (-17.4 to 43.4)	7.0 (-6.5 to 22.4) 6.6 (-3.4 to 17.6) 12.0 (-3.8 to 30.5) 15.9 (-3.5 to 39.1)
PM <sub>2.5</sub>					
Same day Lag 1 day 2-day average 3-day average	-0.5 (-1.3 to 0.3) -0.3 (-1.1 to 0.5) -0.6 (-1.5 to 0.4) -1.1 (-3.1 to 0.9)	-1.9 (-3.5 to -0.3)* -1.2 (-2.8 to 0.3) -2.0 (-3.8 to -0.2)* -3.3 (-7.2 to 0.8)	5.3 (-3.6 to 15) 1.7 (-6.3 to 10.4) 4.3 (-5.4 to 15.1) -17.3 (-33.5 to 2.9)	16.9 (2.2 to 33.6)* 14.6 (0.8 to 30.4)* 22.0 (4.8 to 42.1)* 69.1 (20.1 to 138.2)*	5.1 (-3.6 to 14.5) -3.8 (-12.1 to 5.3) 0.1 (-9.8 to 11.1) 5.8 (-15.8 to 33.0)

2-day average: the median concentration of a pollutant collected on the day and 1 day before the clinical testing was done. 3-day average: the median concentration of a pollutant collected on the day and 2 days before the clinical testing was done. Models were adjusted for testing period, daily temperature, relative humidity, and use of SABA and ICS. Interaction terms were included in the models to test modifying effects of taking ICS (a binary indicator) and sex (binary indicator) on air pollutants.

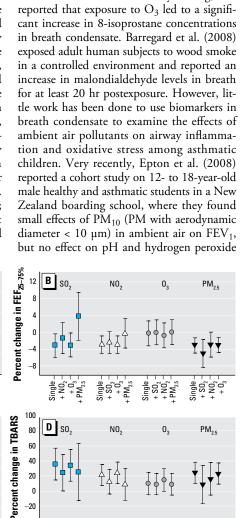
\*p < 0.05.

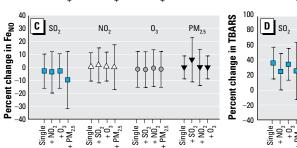
suggesting that the results of each pollutant were independent of other pollutants.

#### **Discussion**

Principal findings of this study are that SO<sub>2</sub>, NO<sub>2</sub>, and PM<sub>2.5</sub> in ambient air were significantly associated with decrements in FEV<sub>1</sub> and FEF<sub>25–75%</sub>, and increases in TBARS in breath condensate among children with asthma. The risk estimates were larger for pollutant concentrations averaged over 3 days than over a shorter term. Clinically FEF<sub>25-75%</sub> has been considered to be sensitive in detecting small airway dysfunction (Lebecque et al. 1993). These associations were robust after inclusion of daily temperature, relative humidity, study period, and the use of asthma medications SABA and ICS in models. The associations for O<sub>3</sub> and PM<sub>2.5</sub> were not sensitive to inclusion of co-pollutants, as adjusting for other pollutants in the model did not markedly change the strength of the risk estimates for O3 and PM<sub>2.5</sub>. The associations between pulmonary function decrements and SO2 and NO2 were sensitive to the inclusion of PM<sub>2.5</sub> in models, suggesting that PM<sub>2.5</sub> may have confounded some of the effects of SO<sub>2</sub> and NO<sub>2</sub>. Adverse changes in FEF<sub>25-75%</sub> were consistent with those of TBARS in response to SO2, NO2, and PM<sub>2.5</sub> with a similar lag structure, suggesting a coherent outcome for small airway function and oxidative stress. Our results on pulmonary function changes were similar to those published previously (Delfino et al. 2004; Epton et al. 2008; Gielen et al. 1997; Romieu et al. 2008), although air pollutant levels in the present study were low compared to those studies.

Exhaled breath condensate has been used in clinical settings to study pathological mechanisms of respiratory illness such as asthma, chronic obstructive pulmonary disease, and cystic fibrosis (Baraldi et al. 2003; Horvath et al. 2005). Elevated concentrations of oxidant nitrate (Balint et al. 2001), IL-6 and leukotriene B4 (Carpagnano et al. 2003), and 8-isoprostane (Montuschi et al. 2000), have been found in breath condensate of adult smokers. Markers of inflammation and oxidative stress in breath were associated with occupational exposure to cobalt and tungsten (Goldoni et al. 2004), chromium (Caglieri et al. 2006), silica (Pelclová et al. 2007), and asbestos (Lehtonen et al. 2007). Montuschi et al. (2002) exposed healthy nonsmoking adults to high concentration of O<sub>3</sub> (400 ppb) and clean air for 2 hr on separate occasions in a laboratory setting, and





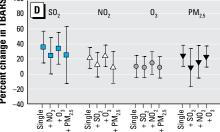


Figure 3. Percent changes and 95% confidence intervals in FEV<sub>1</sub> (A),  $\text{FEF}_{25-75\%}(B)$ ,  $\text{Fe}_{N0}$  (C), and breath TBARS (D) of asthmatic children, in association with an interquartile-range increase in a pollutant, in oneand two-pollutant models. Pollutant concentrations were averaged over 3 days including the same day and 2 days before the clinical testing was done. Models were adjusted for testing period, daily temperature, relative humidity, and use of SABA and ICS.

in exhaled breath. However, with a panel of asthmatic children living in Mexico City, Romieu et al. found an association between malondialdehyde in exhaled breath and PM<sub>2.5</sub> and O<sub>3</sub> in ambient air (Romieu et al. 2008). A 14.2-μg/m<sup>3</sup> (interquartile-range) increase in 8-hr moving average PM<sub>2.5</sub> was significantly associated with a 1.12-nmol increase in malondialdehyde, while a 15.9-ppb (interquartile-range) increase in 8-hr moving average O3 was significantly associated with a 1.16-nmol increase in malondialdehyde. Our results are consistent with these findings in that inhaled ambient air pollutants were significantly associated with an increase in an oxidative stress marker in breath condensate.

Oxidative stress-inducing reactive oxygen species are physiologically active mediators that can be induced by air pollutants. PM<sub>2.5</sub>, SO<sub>2</sub>, NO<sub>2</sub>, and O<sub>3</sub> have been demonstrated to cause formation of excessive amount of reactive oxygen species in airways and in the cardiovascular system in experimental animals, leading to tissue inflammation and cell death (Dye et al. 1997; Meng et al. 2003; Persinger et al. 2002). Oxidative stress has been linked to clinical phenotypes such as asthma and atherosclerosis (Chuang et al. 2007; Walter et al. 2004). TBARS are a group of lowmolecular-weight chemicals that are formed during the decomposition of lipid peroxidation products (Halliwell and Chirico 1993), and thus are often used as an index of lipid peroxidation and oxidative stress (Kodavanti et al. 2000; Rhoden et al. 2004; Walter et al. 2004). 8-Isoprostane is a prostaglandin F2α– like compound produced by free radicals via the nonenzymatic peroxidation of arachidonic acid in membrane phospholipids (Morrow et al. 1990). Because TBARS are derived from oxidative degradation of a broader range of biological substances including lipids and sugars than is 8-isoprostane (Halliwell and Gutteridge 1981; Janero 1990), and because we observed an increase in TBARS but not 8-isoprostane in breath samples in relation to elevated air pollutants, it appears that TBARS may be a more sensitive biomarker than 8-isoprostane in breath condensate as a useful tool for investigating air pollution-related oxidative stress among asthmatic children, especially for acute exposures. It remains to be seen whether immunoassays with significantly higher sensitivity and specificity can improve the efficacy of 8-isoprostane as a biomarker of oxidative stress in exhaled breath condensate.

ICS are thought to reduce inflammation and oxidative stress in airways (Baraldi et al. 2003; Khor et al. 2007). Children with asthma exacerbation have reduced 8-isoprostane in breath condensate after ICS treatment (Baraldi et al. 2003). In the present study, we found that ICS seems to be able to reduce some of the adverse effects of air pollutants,

Percent change in FEV<sub>1</sub>

as the risk estimates appear to be larger for the children who did not take ICS during the study period. Consistent with our findings, previous studies have also reported stronger associations between exposure to air pollutants and decrements in pulmonary function and increases in Fe<sub>NO</sub> for children not taking anti-inflammatory medications than for children taking the medications (Delfino et al. 2006, 2008; Mar et al. 2005). In the present study only 37% of children used ICS, which could result in a reduced statistical power to detect associations with pollutants in this group.

Oxidative stress often occurs along with tissue inflammation, which induces the release of reactive oxygen species to cause tissue injury (Chuang et al. 2007; Dye et al. 1997; Kodavanti et al. 2000). Unlike previously published studies (Delfino et al. 2006; Mar et al. 2005), in our study we did not find statistically significant adverse changes in inflammatory marker FeNO associated with air pollution, and the level of IL-6 in breath condensate was too low to be detectable. One interpretation for the results of inflammation markers was that we might need a larger sample size to detect significant changes in airway inflammation. Another possible explanation might be that the inflammation in lower airways of the children was not captured by Fe<sub>NO</sub> measured at a flow rate of 0.05 L/ sec; Barregard et al. (2008) reported that exposure to wood smoke particles resulted in significant increase in Fe<sub>NO</sub> measured at a flow rate of 0.27 L/sec, but not at 0.05 L/sec. Alternatively, this group of asthmatic children might have been in such a state of airway inflammation that it overwhelmed the effects of air pollution, particularly when exposure concentrations were lower than those in other studies (Delfino et al. 2006; Mar et al. 2005).

We did not find a consistent adverse effect of  $O_3$ .  $O_3$  exhibited a significantly negative association with  $Fe_{NO}$  (Table 4), but a positive association with TBARS (Figure 3). The negative association between  $O_3$  concentration and  $Fe_{NO}$  is counterintuitive, because laboratory studies have shown that  $O_3$  at high concentrations (200–400 ppb) caused inflammation in the airways of human subjects (Balmes et al. 1996; Frampton et al. 1997). In the present study, adjusting for co-pollutants and study periods in the models did not change the results. A sound interpretation for this result remains to be found at this point in time.

When a large number of regression models are run on a data set, some statistically significant associations may occur by chance. To reduce the possibility of spurious relationships due to multiple comparisons, we limited sensitivity analyses to those that consistently demonstrated statistical significance in base models, and considered only results showing a consistent pattern as actual effects.

#### Conclusion

Results from our study demonstrate a significant decrement in small airway function and an increase in airway oxidative stress in asthmatic children in association with SO<sub>2</sub>, NO<sub>2</sub>, and PM<sub>2.5</sub> in ambient air. TBARS in breath condensate may be a useful tool for investigating air pollution-related oxidative stress among asthmatic children. ICS seems to be able to reduce some of the adverse respiratory response to air pollutants.

### REFERENCES

- American Thoracic Society. 1995. Standardization of spirometry: 1994 update. Am J Respir Crit Care Med 152:1107–1136.
- American Thoracic Society. 2005. ATS/ERS recommendations for standardized procedures for the online and offline measurement of exhaled lower respiratory nitric oxide and nasal nitric oxide. Am J Respir Crit Care Med 171:912–930.
- Balint B, Donnelly LE, Hanazawa T, Kharitonov SA, Barnes PJ. 2001. Increased nitric oxide metabolites in exhaled breath condensate after exposure to tobacco smoke. Thorax 56:456–461.
- Balmes JR, Chen LL, Scannell C, Tager I, Christian D, Hearne PQ, et al. 1996. Ozone-induced decrements in  $FEV_1$  and FVC do not correlate with measures of inflammation. Am J Respir Crit Care Med 153:904–909.
- Baraldi E, Carraro S, Alinovi R, Pesci A, Ghiro L, Piacentini G, et al. 2003. Cysteinyl leukotrienes and 8-isoprostane in exhaled breath condensate of children with asthma exacerbations. Thorax 58:505–509.
- Barregard L, Sallsten G, Andersson L, Almstrand A-C, Gustafson P, Andersson M, et al. 2008. Experimental exposure to wood smoke: effects on airway inflammation and oxidative stress. Occup Environ Med 65:319–324.
- Brand HS, Bruins ML, Veerman ECI, Nieuw Amerongen AV. 2006. Secretion rate and amylase concentration of whole saliva after consumption of beer. Int J Dent Hygiene 4:160–161.
- Brauer M, Hoek G, van Vliet P, Meliefste K, Fischer PH, Wijga A, et al. 2002. Air pollution from traffic and the development of respiratory infections and asthmatic and allergic symptoms in children. Am J Respir Crit Care Med 166:1092–1098.
- Burnett RT, Smith-Doiron M, Stieb D, Çakmak S, Brook JR. 1999. Effects of particulate and gaseous air pollution on cardiorespiratory hospitalizations. Arch Environ Health 54:130–139.
- Caglieri A, Goldoni M, Acampa O, Andreoli R, Vettori MV, Corradi M, et al. 2006. The effect of inhaled chromium on different exhaled breath condensates biomarkers among chrome-plate workers. Environ Health Perspect 114:542-546.
- Canadian Institute for Health Information. 2001. Respiratory
  Disease in Canada. Ottawa, Ontario, Canada:Canadian
  Lung Association, Health Canada, Statistics Canada.
  Available: http://www.phac-aspc.gc.ca/publicat/rdc-mrc01/pdf/rdc0901e.pdf [accessed 6 March 2009].
- Carpagnano GE, Kharitonov SA, Foschino-Barbaro MP, Resta O, Gramiccioni E, Barnes PJ. 2003. Increased inflammatory markers in the exhaled breath condensate of cigarette smokers. Eur Respir J 21:589-591.
- Chuang K-J, Chan C-C, Su T-C, Lee C-T, Tang C-S. 2007.

  The effect of urban air pollution on inflammation, oxidative stress, coagulation, and autonomic dysfunction in young adults. Am J Respir Crit Care Med 176:370-376.
- Delfino RJ, Quintana PJE, Floro J, Gastañaga VM, Samimi B, Kleinman MT, et al. 2004. Association of FEV<sub>1</sub> in asthmatic children with personal and microenvironmental exposure to airborne particulate matter. Environ Health Perspect 112:932–941.
- Delfino RJ, Staimer N, Gillen D, Tjoa T, Sioutas C, Fung K, et al. 2006. Personal and ambient air pollution is associated with increased exhaled nitric oxide in children with asthma. Environ Health Perspect 114:1736–1743.
- Delfino RJ, Staimer N, Tjoa T, Gillen D, Kleinman MT, Sioutas C, et al. 2008. Personal and ambient air pollution exposures and lung function decrements in children with asthma. Environ Health Perspect 116:550–558.

- Dye JA, Adler KB, Richards JH, Dreher KL. 1997. Epithelial injury induced by exposure to residual oil fly-ash particles: role of reactive oxygen species. Am J Respir Cell Mol Biol 17:625–633.
- Epton MJ, Dawson RD, Brooks WM, Kingham S, Aberkane T, Cavanagh JA, et al. 2008. The effect of ambient air pollution on respiratory health of school children: a panel study. Environ Health 7:16; doi: 10.1186/1476-069X-7-16 [Online 14 May 2008].
- Frampton MW, Balmes JR, Cox C, Krein PM, Speers DM, Tsai Y, et al. 1997. Effects of ozone on normal and potentially sensitive human subjects. Part III: Mediators of inflammation in bronchoalveolar lavage fluid from nonsmokers, smokers, and asthmatic subjects exposed to ozone: a collaborative study. Res Rep Health Eff Inst 78:73–79.
- Gielen MH, van der Zee SC, van Wijnen JH, van Steen CJ, Brunekreef B. 1997. Acute effects of summer air pollution on respiratory health of asthmatic children. Am J Respir Crit Care Med 155:2105–2108.
- Goldoni M, Catallani S, De Palma G, Manini P, Acampa O, Corradi M, et al. 2004. Exhaled breath condensate as a suitable matrix to assess lung dose and effects in workers exposed to cobalt and tungsten. Environ Health Perspect 112:1293–1298.
- Halliwell B, Chirico S. 1993. Lipid peroxidation: its mechanism, measurement, and significance. Am J Clin Nutr 57(suppl):715S-725S.
- Halliwell B, Gutteridge JMC. 1981. Formation of thiobarbituricacid-reactive substance from deoxyribose in the presence of iron salts: the role of superoxide and hydroxyl radicals. FEBS Lett 128:347–352.
- Halliwell B, Gutteridge JMC. 1989. Lipid peroxidation: a radical chain reaction. In: Free Radicals in Biology and Medicine (Halliwell B, Gutteridge JMC, eds.). Oxford, UK:Clarendon Press. 188–249.
- Hankinson JL, Odencrantz JR, Fedan KB. 1999. Spirometric reference values from a sample of the general U.S. population. Am J Respir Crit Care Med 159:179–187.
- Horvath I, Hunt J, Barnes PJ. 2005. Exhaled breath condensate: methodological recommendations and unresolved questions. Eur Respir J 26:523–548.
- Hulsmann AR, Raatgeep HR, den Hollander JC, Stijnen T, Saxena PR, Kerrebijn KF, et al. 2007. Oxidative epithelial damage produces hyperresponsiveness of human peripheral airways. Am J Respir Crit Care Med 149:519–525.
- Janero DR. 1990. Malondialdehyde and thiobarbituric acidreactivity as diagnostic indices of lipid peroxidation and peroxidative tissue injury. Free Rad Biol Med 9:515–540.
- Khor YH, Feltis BN, Reid DW, Ward C, Johns DP, Wood-Baker R, et al. 2007. Airway cell and cytokine changes in early asthma deterioration after inhaled corticosteroid reduction. Clinical and Experimental Allergy 37:1189–1198.
- Kodavanti UP, Schladweiler MC, Ledbetter AD, Watkinson WP, Campen MJ, Winsett DW, et al. 2000. The spontaneously hypertensive rat as a model of human cardiovascular disease: evidence of exacerbated cardiopulmonary injury and oxidative stress from inhaled emission particulate matter. Toxicol Appl Pharmacol 164:250–263.
- Krahn MD, Berka C, Langlois P, Detsky AS. 1996. Direct and indirect costs of asthma in Canada, 1990. CMAJ 154:821–831.
- Lebecque P, Kiakulanda P, Coates AL. 1993. Spirometry in the asthmatic child: is FEF<sub>25-75</sub> a more sensitive test than FEV1/FVC? Pediatr Pulmonol 16:19–22.
- Lehtonen H, Oksa P, Lehtimäki L, Sepponen A, Nieminen R, Kankaanranta H, et al. 2007. Increased alveolar nitric oxide concentration and high levels of leukotriene B<sub>4</sub> and 8-isoprostane in exhaled breath condensate in patients with asbestosis. Thorax 62:602-607.
- Mar TF, Jansen K, Shepherd K, Lumley T, Larson TV, Koenig JQ. 2005. Exhaled nitric oxide in children with asthma and short-term PM<sub>2.5</sub> exposure in Seattle. Environ Health Perspect 113:1791–1794.
- Meng Z, Qin G, Zhang B, Geng H, Bai Q, Bai W, et al. 2003. Oxidative damage of sulfur dioxide inhalation on lungs and hearts of mice. Environ Res 93:285–292.
- Montuschi P. 2005. New Perspectives in Monitoring Lung Inflammation: Analysis of Exhaled Breath Condensate. Boca Raton, FL:CRC Press.
- Montuschi P, Collins JV, Ciabattoni G, Lazzeri N, Corradi M, Kharitonov SA, et al. 2000. Exhaled 8-Isoprostane as an in vivo biomarker of lung oxidative stress in patients with COPD and healthy smokers. Am J Respir Crit Care Med 162:1175–1177.
- Montuschi P, Nightingale JA, Kharitonov SA, Barnes PJ. 2002.

- Ozone-induced increase in exhaled 8-isoprostane in healthy subjects is resistant to inhaled budesonide. Free Rad Biol Med 33:1403–1408.
- Montuschi P, Ragazzoni E, Valente S, Corbo G, Mondino C, Ciappi G, et al. 2003. Validation of 8-isoprostane and prostaglandin  $\rm E_2$  measurements in exhaled breath condensate. Inflamm Res 52:502–506.
- Morrow JD, Hill KE, Burk RF, Nammour TM, Badr KF, Roberts LJ. 1990. A series of prostaglandin F2-like compounds are produced *in vivo* in humans by a non-cyclooxygenase, free radical-catalyzed mechanism. Proc Natl Acad Sci USA 87:9383–9387.
- Pelclová D, Fenclová Z, Kacer P, Navrátil T, Kuzma M, Lebedová JK, et al. 2007. 8-Isoprostane and leukotriene in exhaled breath condensate in Czech subjects with silicosis. Industrial Health 49:766–774.
- Persinger RL, Poynter ME, Ckless K, Janssen-Heininger YM. 2002. Molecular mechanisms of nitrogen dioxide induced epithelial injury in the lung. Molecular Cell Biochem 234–235:71–80.

- Rhoden CR, Lawrence J, Godleski JJ, González-Flecha B. 2004. N-acetylcysteine prevents lung inflammation after short-term inhalation exposure to concentrated ambient particles. Toxicol Sci 79:296–303.
- Romieu I, Barraza-Villarreal A, Escamilla-Nuñez C, Almstrand AC, Diaz-Sanchez D, Sly PD, et al. 2008. Exhaled breath malondialdehyde as a marker of effect of exposure to air pollution in children with asthma. J Allergy Clin Immunol 121:930–939.
- Stieb DM, Beveridge RC, Brook JR, Smith-Doiron M, Burnett RT, Dales RE, et al. 2000. Air pollution, aeroallergens and cardiorespiratory emergency department visits in Saint John, Canada. J Expo Anal Environ Epidemiol 10:461–477.
- Sunyer J, Spix C, Quénel P, Ponce-de-Deón A, Pönka A, Barumandzadeh T, et al. 1997. Urban air pollution and emergency admissions for asthma in four European cities: the APHEA Project. Thorax 52:760–765.
- Vichit-Vadakan N, Ostro BD, Chestnut LG, Mills DM, Aekplakorn W, Wangwongwatana S, et al. 2001. Air pollution and respiratory symptoms: results from three panel studies

- in Bangkok, Thailand. Environ Health Perspect 109(suppl 3):381–387.
- Villeneuve PJ, Chen L, Rowe BH, Coates F. 2007. Outdoor air pollution and emergency department visits for asthma among children and adults: a case-crossover study in northern Alberta, Canada. Environ Health 6:40; doi:10.1186/1476-069X-6-40 [Online 24 December 2007].
- Walter MF, Jacob RF, Jeffers B, Ghadanfar MM, Preston GM, Buch J, et al. 2004. Serum levels of thiobarbituric acid reactive substances predict cardiovascular events in patients with stable coronary artery disease: a longitudinal analysis of the PREVENT study. J Am Coll Cardiol 44:1996–2002.
- Wong C-M, Atkinson RW, Anderson HR, Hedley AJ, Ma S, Chau PYK, et al. 2001. A tale of two cities: effects of air pollution on hospital admissions in Hong Kong and London compared. Environ Health Perspect 110:67–77.
- Yagi K. 1982. Assay for serum lipid peroxide level and its clinical significance. In: Lipidperoxide in Biology and Medicine (Yagi K, ed.). New York:Academic Press, 232–242.